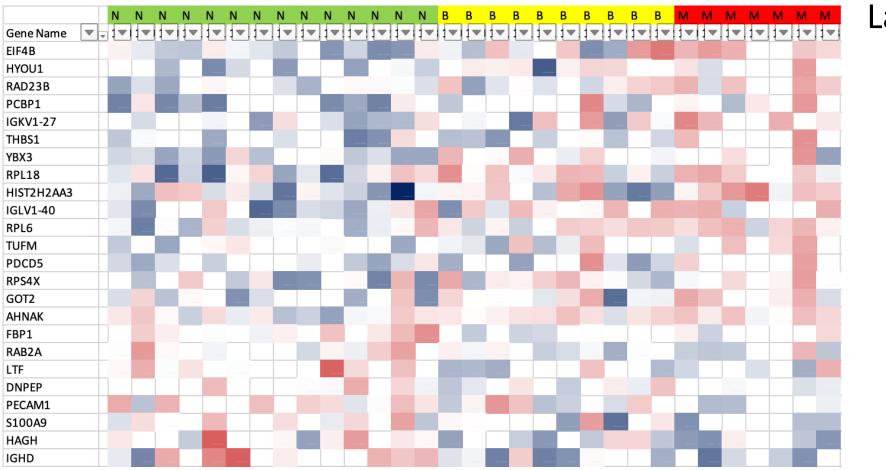
3D Printed Microneedles for Breast Cancer Biomarker Discovery in Dermal Interstitial Fluid

> Andy Hung Mentors: Joseph DeSimone, Sharon Pitteri, Steven Poplack 1/11/2023

#### Two Goals

- 1. Create a microneedle system for reliable painless skin ISF sampling
- 2. Discover breast cancer protein biomarkers in skin ISF

#### A Pilot Study of Breast Cancer ISF Biomarker



Laser micropores



↓ 1 hr vacuum



5 μL ISF

Bermudez, Early Cancer Detection Conference Poster, 2020

#### ISF is a Promising Diagnostic Compartment

#### Table 2 | Comparison of analytes representing several classes found in blood plasma, ISF, saliva and sweat

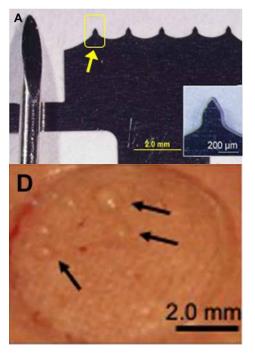
	Na <sup>+</sup>	<b>K</b> +	Lactate	Glucose	Cortisol	Drugs	Cytokines	Antibodies
Molecular weight (Da)	23	39	90	180	362	Mostly hundreds of daltons	More than five to tens of kilodaltons	Hundreds of kilodaltons
Lipophilicity	Very low (charged)	Very low (charged)	Very low (charged)	Low (hydroxyls)	High	Often high	Very low	Very low
Blood plasma	135-145 mM	3.5-5 mM	0.5-10 mM (resting to nonresting)	4.1-6.9 mM (venous, resting)	Hundreds of nanomolar total; tens of nanomolar unbound fraction	Mostly equivalent to unbound in plasma	Picomolar to nanomolar	Varies; total ~0.4-16 mg ml <sup>-1</sup>
ISF <sup>a</sup>	Similar to plasma	Similar to plasma	Similar to plasma	Similar to plasma	Unbound similar to plasma (p)	Many equivalent to unbound in plasma (p)	80% of plasma (a,p)	15-25% of plasma

#### ISF Continuous Glucose Monitoring



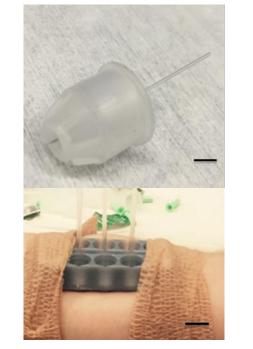
#### **ISF Sampling Strategies**

Microneedling + Vacuum

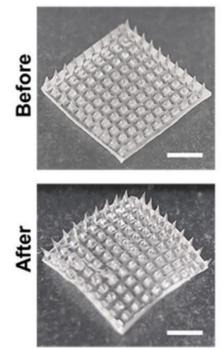


2-5 µL in 20 min

32G Needles + Positive pressure



Hydrogel Microneedles



10 µL in 30 min (64%)

 $4-8\ \mu\text{L}$  in  $3\ min$ 

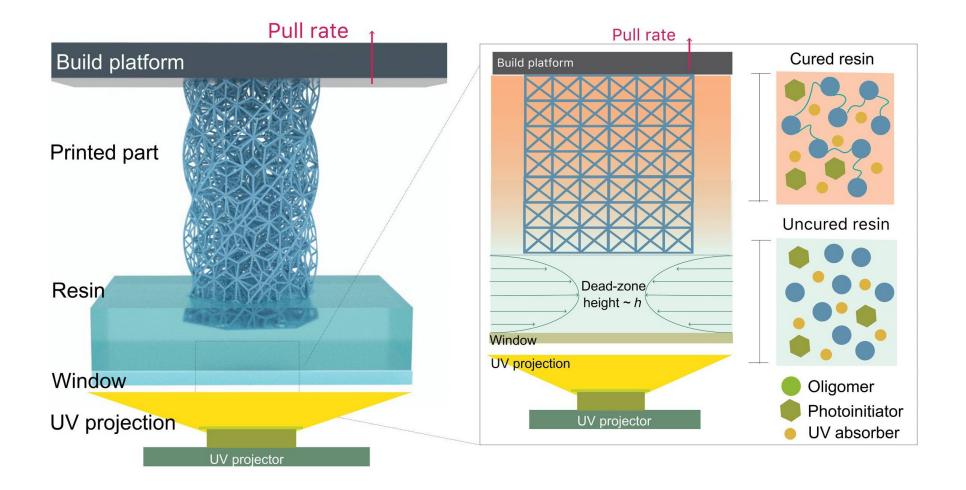


# Can we leverage 3D printing to improve ISF collection?

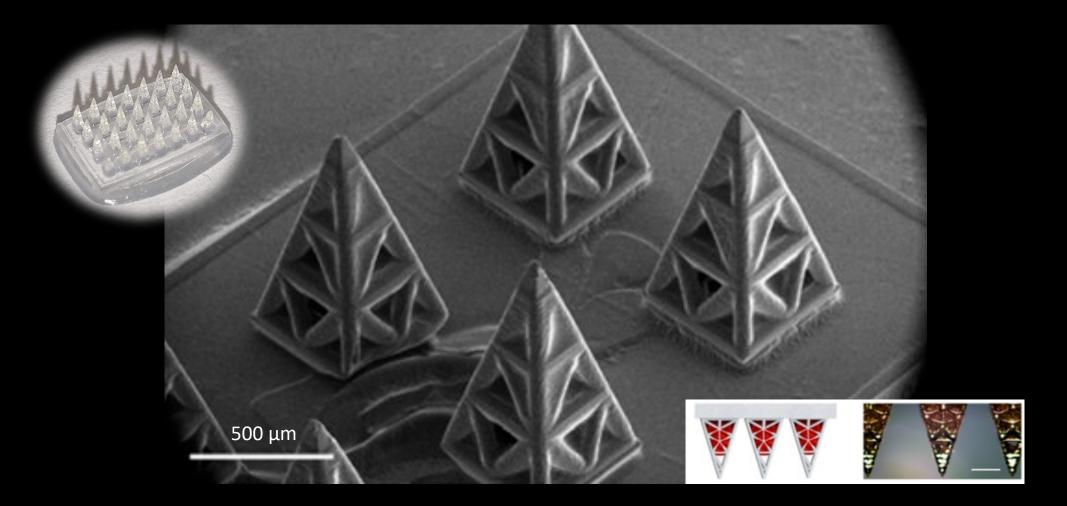
#### Design Criteria

- Collect 50 uL in 20 minutes (max est. 60  $\mu$ L/cm<sup>2</sup>)
- Consistent penetration
- Mechanically robust
- Easy ISF retrieval from device
- Easy to apply
- Can sterilize

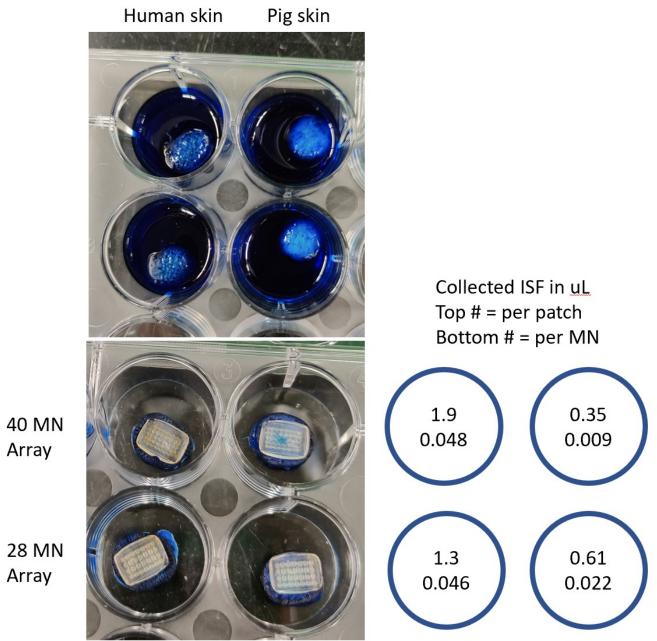
#### Continuous Liquid Interface Production (CLIP)



#### Lattice Microneedle Array Patch (LMAP)



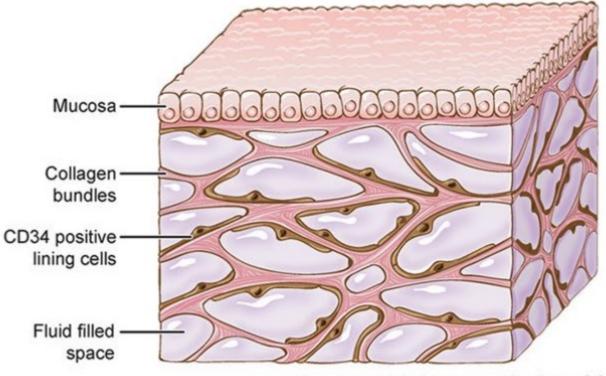
## Does LMAP collect ISF in skin?



40 MN Array

Array

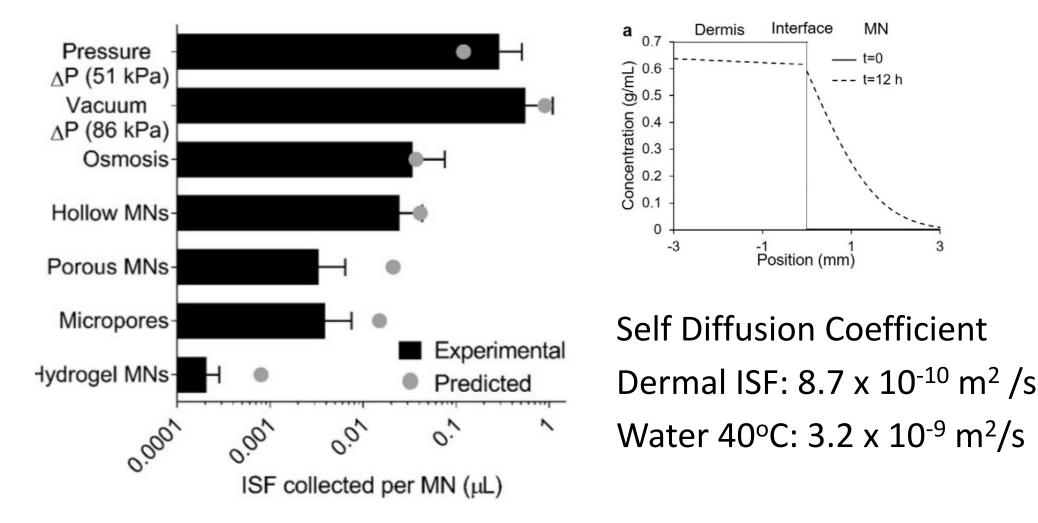
#### Fluid in skin is not free fluid



- Proteoglycans
- Glycoproteins
- Hyaluronic acid

J Gregory ©2016 Mount Sinai Health System

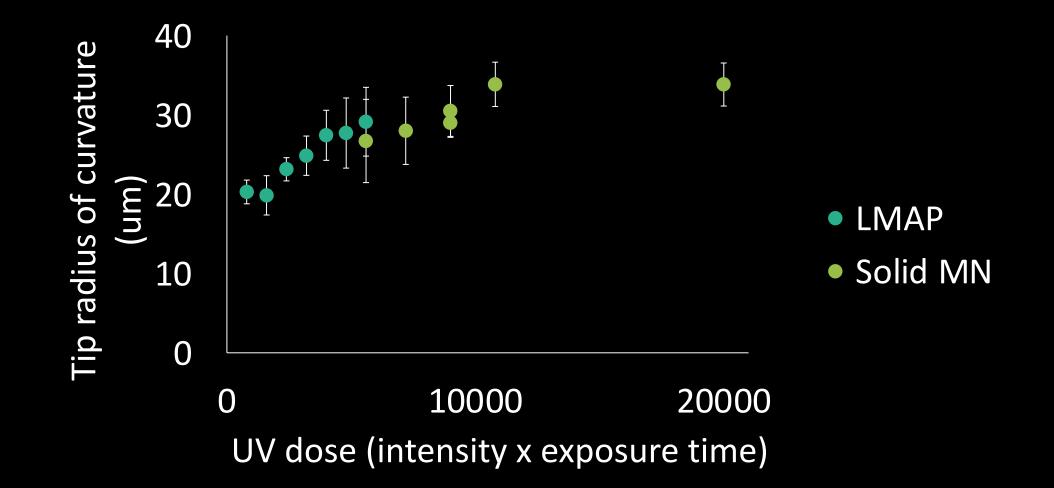
#### Diffusion alone is probably not enough



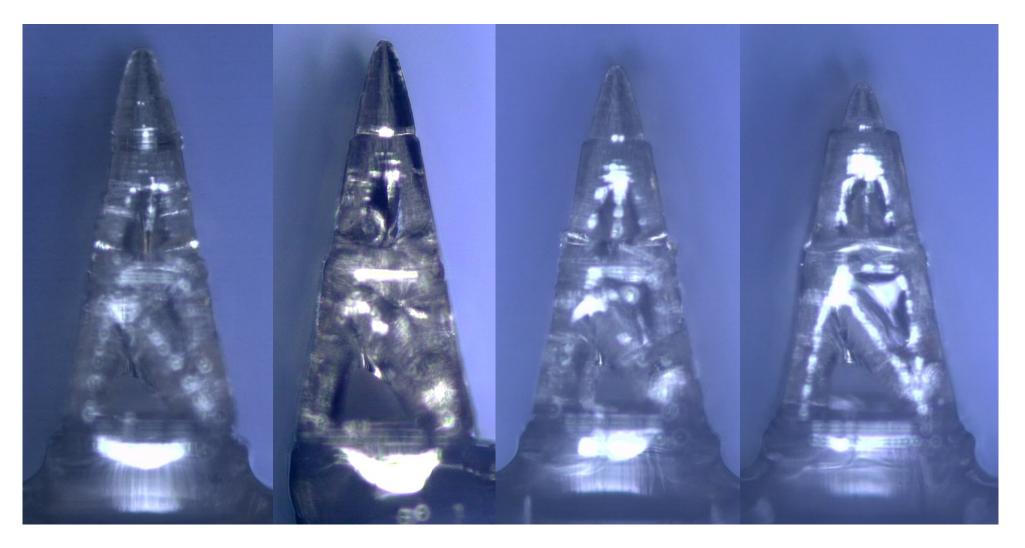
### Design Criteria

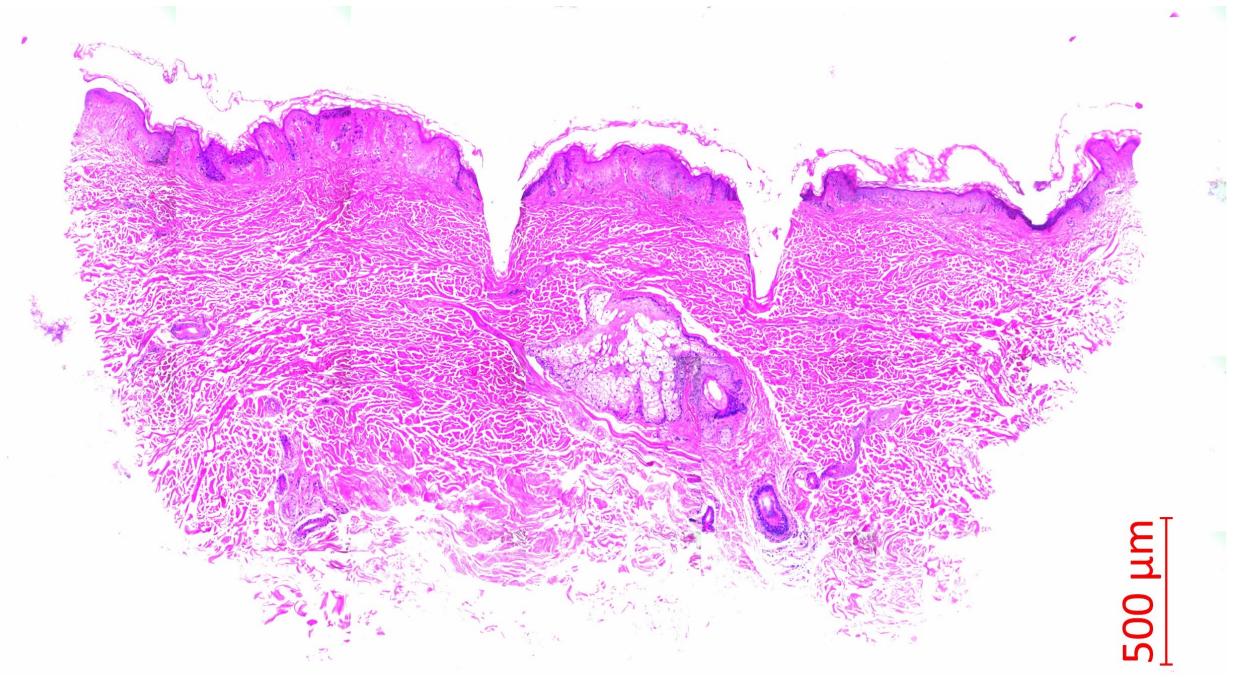
- Collect 50 uL in 20 minutes
- Consistent penetration
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#### Optimize UV dose for tip sharpness



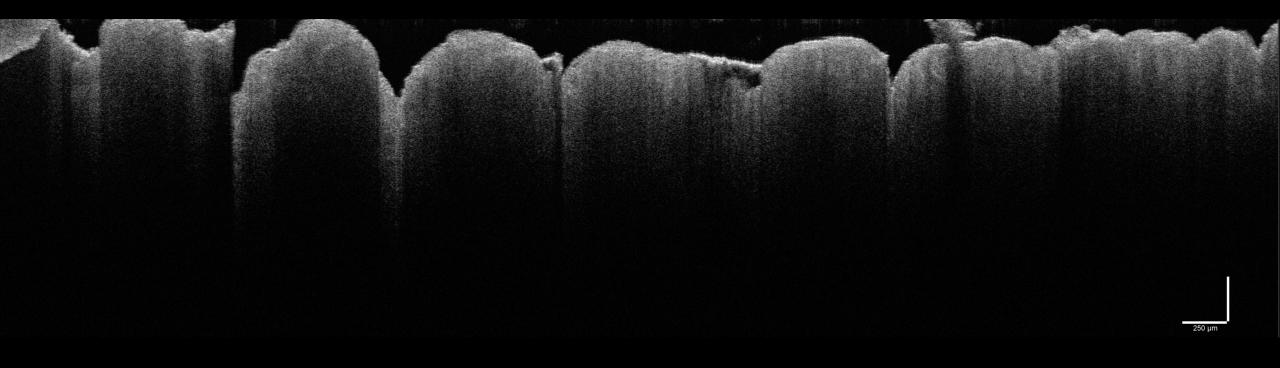
UV Dose (a.u.)	5600	2400	1600	800
Tip Radius (um)	29 ± 4	23 ± 1	20 ± 2	20 ± 1





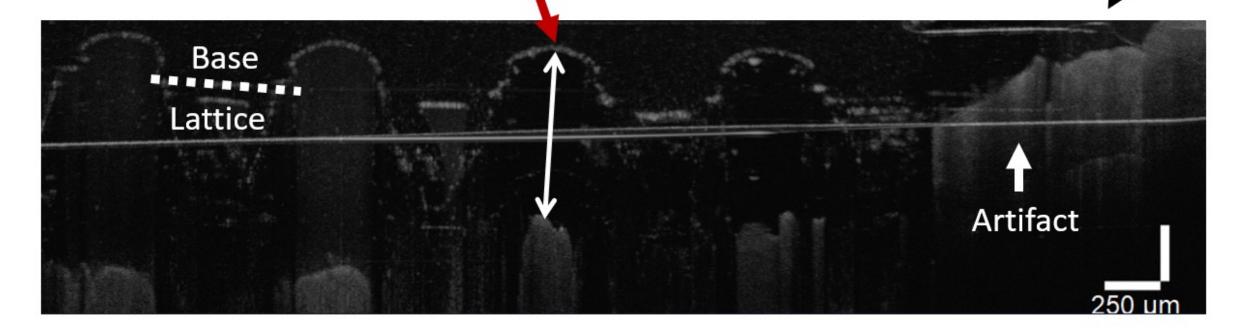
Collaboration with Davis Lab

#### OCT of human skin after LMAP application



#### Use OCT to assess penetration

Back plate to skin distance 800-1050 um (40-21% penetration efficiency)

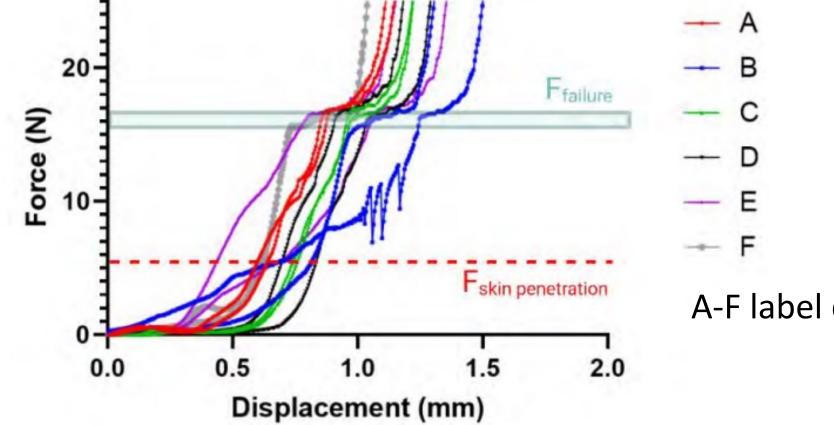


Edge of patch

### Design Criteria

- Collect 50 uL in 20 minutes
- Consistent penetration
- Mechanically robust
- Easy ISF retrieval from device
- Easy to apply
- Can sterilize

#### A Safety Factor of 3 in Axial Loading



A-F label different designs

Courtesy of Netra Rajesh

#### Microneedles bend without fracture



#### Learnings

- Collect 50 uL in 20 minutes: ΔP likely necessary
- Consistent penetration: 20-30  $\mu m$  sharpness ok, 50% of design height penetrates, consistency TBD
- Mechanically robust: axial loading  $\checkmark$ , lateral force evaluation needed
- Easy ISF retrieval from device
- Easy to apply
- Can sterilize

#### Acknowledgement

Mentors: Joseph DeSimone, Sharon Pitteri, Steven Poplack Desimone Lab: Netra Rajesh and Gunilla Jacobson Stanford Cancer Imaging Training Program

