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Computational challenges and opportunities for a bi-directional artificial retina

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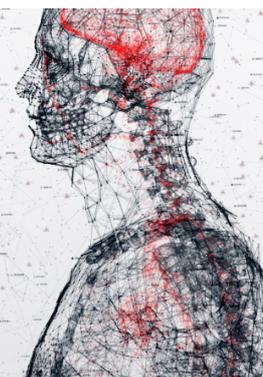
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PAPER

Computational challenges and opportunities for a bi-directional artificial retina

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21 October 2020Nishal P Shah^{1,2,3,5}  and E. J. Chichilnisky^{2,3,4}¹ Department of Electrical Engineering, Stanford University, Stanford, CA, United States of America² Hansen Experimental Physics Laboratory, Stanford University, Stanford, CA, United States of America³ Department of Neurosurgery, Stanford University, Stanford, CA, United States of America⁴ Department of Ophthalmology, Stanford University, Stanford, CA, United States of America⁵ Author to whom any correspondence should be addressed.E-mail: bhaishahster@gmail.com**Keywords:** retinal prosthesis, artificial retina, electrical stimulation, brain-machine interface, neural code, artificial vision**Abstract**

A future artificial retina that can restore high acuity vision in blind people will rely on the capability to both read (observe) and write (control) the spiking activity of neurons using an adaptive, bi-directional and high-resolution device. Although current research is focused on overcoming the technical challenges of building and implanting such a device, exploiting its capabilities to achieve more acute visual perception will also require substantial computational advances. Using high-density large-scale recording and stimulation in the primate retina with an *ex vivo* multi-electrode array lab prototype, we frame several of the major computational problems, and describe current progress and future opportunities in solving them. First, we identify cell types and locations from spontaneous activity in the blind retina, and then efficiently estimate their visual response properties by using a low-dimensional manifold of inter-retina variability learned from a large experimental dataset. Second, we estimate retinal responses to a large collection of relevant electrical stimuli by passing current patterns through an electrode array, spike sorting the resulting recordings and using the results to develop a model of evoked responses. Third, we reproduce the desired responses for a given visual target by temporally dithering a diverse collection of electrical stimuli within the integration time of the visual system. Together, these novel approaches may substantially enhance artificial vision in a next-generation device.

1. Introduction

Neural interfaces are designed to restore or augment human capabilities, by either reading out or writing in the electrical activity of neurons. Read-out interfaces typically use neural activity to decode user intentions, for applications such as limb and cursor movements (Bacher *et al* 2015, Jarosiewicz *et al* 2015, Pandarinath *et al* 2017) or speech (Anumanchipalli *et al* 2019, Stavisky *et al* 2019). Write-in interfaces typically disrupt pathological neural activity for treating neurodegenerative diseases (Lozano and Lipsman 2013) or stimulate sensory neurons to restore audition (Shannon 2012), touch (Tyler 2015) or vision (Goetz and Palanker 2016). Future neural interfaces will likely enable applications such as controlling complex robotic devices, extending memory, or augmenting natural senses with artificial inputs (Marcus and Koch 2014). These applications will

likely require bi-directional capability, i.e. the ability to both read and write electrical activity in neural circuits. For example, read-write capability will be useful in providing sensory feedback about limb motion in motor prostheses, or in optimizing electrical stimulation using the measured ongoing and evoked activity in sensory prostheses. Furthermore, to emulate and harness the natural resolution of signals in the nervous system, future devices will ideally function at single-cell, single-spike resolution. Neural interface functionality with this kind of adaptability and precision will require new approaches for accurate, efficient and fast information processing to match (or even exceed) the natural function of the neural circuit. Hence, important computational challenges must be addressed in order to realize the capabilities of such devices.

In this paper, we describe some of the key computational problems, and approaches to solving them,

for a bi-directional, high-resolution *artificial retina*, which is designed to restore vision in people blinded by retinal degeneration. The retina is a thin layer of neural tissue at the back of the eye that performs the first steps in visual processing (Dowling 1987, Wandell 1995, Rodieck 1998). Light from the visual scene is transduced by the photoreceptors into electrical signals, which are then processed by an extensive network consisting of many types of interneurons, and finally are encoded in the activity of each of ~ 20 functionally distinct retinal ganglion cell (RGC) types that project to diverse targets in the brain (Dacey 2004). In certain conditions, notably age-related macular degeneration and retinitis pigmentosa, photoreceptor degeneration leads to incurable blindness. However, many ganglion cells remain excitable and connected to the brain, raising the possibility of artificially activating them to elicit visual perception. To accomplish this, *retinal prostheses* have been developed to electrically target various parts of the degenerated retina, either epiretinally at the RGC layer (Humayun *et al* 2012), sub-retinally at the interneuron layer (Stingl *et al* 2013, Palanker *et al* 2020) or supra-choroidally outside the retina (Ayton *et al* 2014). Although these devices have been able to restore some degree of vision, the resulting visual sensations are limited to rudimentary shapes such as blobs and arcs, and do not approach the high resolution, and functional value of natural vision (Goetz and Palanker 2016). The performance of these devices is limited in part by the fact that they stimulate the diverse types of retinal neurons coarsely and non-selectively, and thus fail to reproduce the spatially and temporally precise, cell-type specific neural code of the retina.

We propose that a future *artificial retina* will more faithfully restore vision by replicating, as accurately as possible, the precise patterns of firing in the diverse RGC types in response to visual stimuli. To do so, the device will need to (a) use high-resolution electrical recordings of spontaneous activity to identify the cells and cell types in the region of the implant and their expected contribution to visual perception, (b) use electrical stimulation and recording to determine which cell(s) can be stimulated at what current level(s) and by which electrode(s) in the implant, and (c) use electrical stimulation to evoke the optimal achievable pattern of RGC activity for the incoming image, given the measured constraints of the neural interface.

Achieving these goals requires many advances in hardware as well as computation (Muratore and Chichilnisky 2020). Multiple efforts are currently focused on building high-density implantable electrode arrays and interface electronics (Dragas *et al* 2017, Jun *et al* 2017, Muratore *et al* 2019, Obaid *et al* 2020). As these hardware development efforts proceed, we can in parallel tackle the many accompanying computational problems that arise,

using a large-scale high-resolution multi-electrode array recording and stimulation system as an *ex vivo* laboratory prototype of a future implanted device (figure 1(A)). Using this experimental approach (Litke *et al* 2004, Frechette *et al* 2005, Hottowy *et al* 2008, Sekirnjak *et al* 2008, Hottowy *et al* 2012), we identify and target different RGC types in the primate retina by electrical recording coupled with visual and electrical stimulation (figure 1(B)). This approach features small electrodes ($\sim 10 \mu\text{m}$) that are densely packed ($\sim 30\text{--}60 \mu\text{m}$ spacing), and brief (~ 0.1 ms) epiretinal stimulation at low current amplitudes ($\sim 1 \mu\text{A}$) to activate RGCs with single-cell, single-spike resolution (Sekirnjak *et al* 2007, 2008, Jepson *et al* 2013, 2014a). Based on experiments and analysis using this laboratory prototype, we propose that three major classes of computational problems must be solved to produce high-resolution artificial vision:

(1) How does the retina normally represent visual information? Decades of research have focused on developing an accurate *encoding model* $R \sim g(S)$, that can predict the responses (R) of RGCs to any given visual stimulus (S) (figure 1(C)) (Gollisch and Meister 2010). This work has revealed that many different RGC types encode distinct features of the visual stimulus in their distinct spatial and temporal patterns of activity, providing a gold standard of performance for any artificial retina (figure 1(B)). More recently, some studies have also developed *decoding models* $S \sim h(R)$, to estimate the visual stimulus from recorded RGC responses, as a proxy for understanding how downstream neural circuitry in the brain could process the retinal signal (Rieke 1996, Warland *et al* 1997, Stanley *et al* 1999). These two modeling approaches have been largely separate in the computational neuroscience literature. Below we will argue that they find a meaningful meeting point in the neuroengineering application (Nirenberg and Pandarinath 2012). Therefore, in what follows, we summarize encoding and decoding with a single utility function, $f(S, R)$ (figure 1(C)), which measures the degree of correspondence between a visual stimulus (S) and RGC firing (R).

(2) How does the retina respond to electrical stimulation? In principle, if we were able to selectively activate every target cell while avoiding other cells, arbitrary firing patterns in the diverse RGC types could be reproduced. However, existing devices instead stimulate many RGCs of all types simultaneously and non-selectively, providing only coarse control over neural activity and failing to provide the asynchronous, distinctive visual signals normally carried by the diverse RGC types to the brain. Although future high-resolution electrode arrays will allow more precise stimulation, perfect cellular selectivity will remain a challenge. Therefore, we present an approach to determine how to use the available electrical stimuli as effectively as possible in an implanted

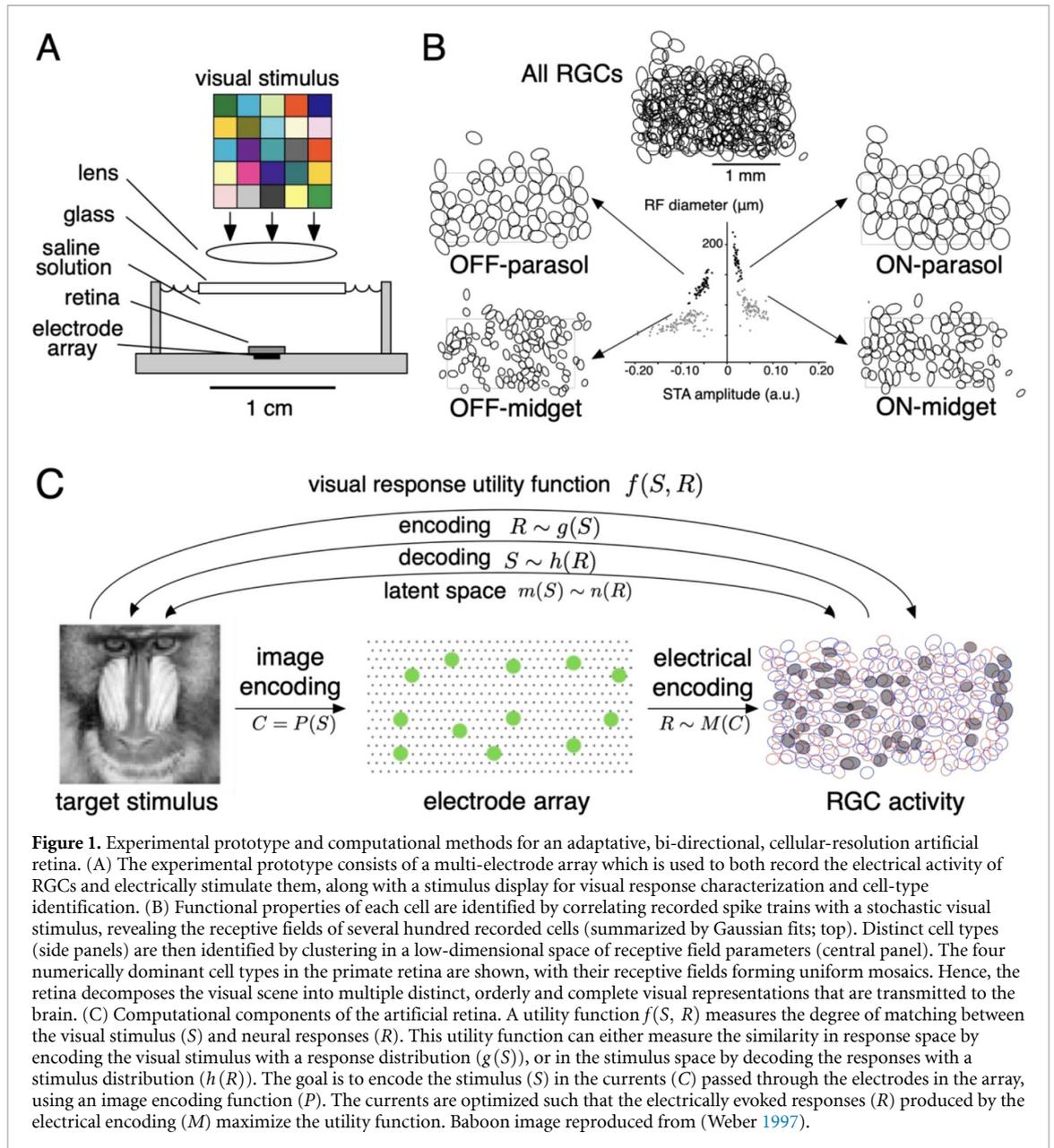


Figure 1. Experimental prototype and computational methods for an adaptive, bi-directional, cellular-resolution artificial retina. (A) The experimental prototype consists of a multi-electrode array which is used to both record the electrical activity of RGCs and electrically stimulate them, along with a stimulus display for visual response characterization and cell-type identification. (B) Functional properties of each cell are identified by correlating recorded spike trains with a stochastic visual stimulus, revealing the receptive fields of several hundred recorded cells (summarized by Gaussian fits; top). Distinct cell types (side panels) are then identified by clustering in a low-dimensional space of receptive field parameters (central panel). The four numerically dominant cell types in the primate retina are shown, with their receptive fields forming uniform mosaics. Hence, the retina decomposes the visual scene into multiple distinct, orderly and complete visual representations that are transmitted to the brain. (C) Computational components of the artificial retina. A utility function $f(S, R)$ measures the degree of matching between the visual stimulus (S) and neural responses (R). This utility function can either measure the similarity in response space by encoding the visual stimulus with a response distribution ($g(S)$), or in the stimulus space by decoding the responses with a stimulus distribution ($h(R)$). The goal is to encode the stimulus (S) in the currents (C) passed through the electrodes in the array, using an image encoding function (P). The currents are optimized such that the electrically evoked responses (R) produced by the electrical encoding (M) maximize the utility function. Baboon image reproduced from (Weber 1997).

device. In what follows, we summarize the RGC responses (R) evoked by current passed through the electrodes (C) with an electrical response model (M), $R \sim M(C)$ (figure 1(C)).

(3) How do we optimally replicate RGC firing in a blind retina? Previous studies have proposed to optimize visual encoding while assuming high selectivity of stimulation (Nirenberg and Pandarinath 2012) or while measuring selectivity using the elicited visual sensation reported by a blind person (Beyeler *et al* 2019). In contrast, we use single-cell resolution measurements of the electrical response properties of RGCs to convert a target visual stimulus (S) into an electrical stimulus (C) in real time. This is accomplished by identifying the electrical encoding of the incident image (P) that maximizes the expected visual utility function (f) between the target visual stimulus (S) and the probabilistic responses (R) generated via

the electrical response function (M).

$$C = P(S) = \operatorname{argmax} \mathbb{E}_{R \sim M(C)} f(S, R) \quad (1)$$

This paper presents a detailed description of the above computational problems, progress towards addressing them, and possible future directions. Then, system design issues are discussed for implementing the solutions in distributed hardware. Finally, the caveats of the approach, and potential application of similar ideas in other neural interfaces, are considered.

2. How does the retina normally represent visual information?

In this section, we focus on identifying the visual response properties of RGCs, captured by a utility function $f(S, R)$ which measures how closely the

visual stimulus S and neural responses R correspond. Standard methods for identifying $f(\cdot)$ are based on recording responses to visual stimuli, which is not possible in a blind retina. Instead, we first use light-evoked recordings from healthy retinas to model $f(\cdot)$, and then adapt the information for a blind retina, in which only recordings of spontaneous activity are available. A major difficulty that arises is the variability of visual responses across different healthy retinas. We describe a method to systematically characterize this inter-retina variability and then to identify the optimal utility function in a blind person.

2.1. Identifying the utility function in a healthy retina

The utility function is most frequently examined in the response space, by transforming the visual stimulus into predicted responses, and then comparing this prediction to measured light responses in the retina. Specifically, the utility function is expressed as

$$f(S, R) = \rho(g(S), R) \quad (2)$$

where the encoder $g(\cdot)$ produces a predicted neural response (or a distribution over possible responses) based on the visual stimulus, and $\rho(\cdot)$ is a measure of similarity between responses (or distributions). Often, $\rho(\cdot)$ is given as the negative log-likelihood of the data given the probabilistic prediction $g(\cdot)$. Many previous studies have modeled $g(\cdot)$ using a cascade of linear and nonlinear processing stages (see (Chichilnisky 2001)), with extensions to include coupling between nearby cells (Mastrorarde 1983, Pillow *et al* 2008, Greschner *et al* 2011), adaptation (Smirnakis *et al* 1997, Chander and Chichilnisky 2001, Kim and Rieke 2001, Carandini and Heeger 2011, Kastner and Baccus 2013), or spatial nonlinearity (Hochstein and Shapley 1976, Demb *et al* 1999, 2001, Freeman *et al* 2015, Liu *et al* 2017, Shah *et al* 2020). Recent studies have also used machine learning to improve encoding models (Batty *et al* 2016, McIntosh *et al* 2016). Finally, instead of using negative log-likelihood or mean-squared error to measure the accuracy of model predictions, some studies have proposed similarity measures $\rho(\cdot)$ that account for response variations unrelated to the stimulus (Victor and Purpura 1996, 1997, van Rossum 2001, Victor 2005, Dubbs *et al* 2010, Ganmor *et al* 2015). Recently, machine learning methods have been developed to estimate a response similarity measure by minimizing the distance between pairs of responses generated by the same stimulus relative to the distance between pairs of responses generated by different stimuli (Shah *et al* 2018).

A complementary approach is to measure similarity in the visual stimulus space, instead of the response space, by modeling how the brain could decode the visual stimulus from retinal responses (Rieke 1996).

In this case, the loss function is expressed as

$$f(S, R) = \sigma(S, h(R)) \quad (3)$$

where the decoder $h(\cdot)$ estimates the visual stimulus (or a distribution over possible visual stimuli) from neural responses and $\sigma(\cdot)$ is a measure of perceptual similarity between visual stimuli (or distributions of stimuli). Since the neural decoding function implemented by the brain is unknown, simplifying assumptions are required. A reasonable assumption is that the brain optimally decodes the neural responses to estimate the visual stimulus, based on the statistics of retinal responses to natural stimuli. A simple version of this idea is a linear stimulus decoder (Warland *et al* 1997, Stanley *et al* 1999, Golden *et al* 2019, Shah *et al* 2019a) selected to minimize the mean-squared error between the true and decoded stimulus. Examples of more elaborate nonlinear decoders include explicitly inverting the encoding models using priors from natural images (Pillow *et al* 2008, Brackbill *et al* 2017), or using supervised machine learning (Parthasarathy *et al* 2017, Zhang *et al* 2020). Similarly, more accurate measures of decoding error could be used, based on perceptual error metrics that capture the features of images that humans can most readily discriminate (Wang *et al* 2004). An advantage of the decoding approach is that, in principle, it allows performance to be measured in the domain of greatest interest: the similarity of the perceived and incident visual image (or specific aspects of them).

A third alternative is to represent both the visual stimulus and neural responses in an intermediate latent space in which similarity can be measured with a simple function. In this formulation,

$$f(S, R) = \|m(S) - n(R)\|_2 \quad (4)$$

where the visual stimulus is transformed to the latent space by $m(\cdot)$, neural responses are transformed to the latent space by $n(\cdot)$, and $\|\cdot\|_2$ is the mean-squared error cost. This formulation is useful when the similarity in either response or stimulus space is unknown, but many visually evoked responses are available to estimate both m and n . For example, this approach might be useful when we want to identify a utility function for specific tasks, such as navigation or reading, for which the standard methods of measuring similarity in visual stimulus space are too general. This approach may also be suitable for other sensory modalities, such as olfaction, in which it is difficult to measure the similarity between different stimuli (Su *et al* 2009).

Irrespective of the exact implementation of the utility function $f(\cdot)$, the differences between visual stimulus properties of different healthy retinas recorded using the experimental lab prototype, and future application in the retina of a blind person, must be considered. Most experimental studies on visual encoding in the retina have either used simple white

noise stimuli, or highly structured stimuli, such as moving bars or gratings, that isolate specific computational mechanisms. However, in natural viewing, the statistics of stimuli are very different, and the eyes are constantly saccading to different parts of the visual scene, with microsaccades and jitter around the fixation location (Kowler 2011). Hence a population of RGCs in a region of the retina sees natural textures and edges through a tiny moving window. Recently, some studies have begun to develop computational models to explain RGC responses to such stimuli (Segal *et al* 2015, Batty *et al* 2016, Heitman *et al* 2016, McIntosh *et al* 2016, Salisbury and Palmer 2016, Turner and Rieke 2016, Maheswaranathan *et al* 2019, Turner *et al* 2019, Shah *et al* 2020). Visual stimuli can also be specialized to tasks that are important for a blind person, such as indoor navigation or reading, which probably involve different stimulus statistics.

In addition to the visual stimulus, differences in the distribution of the visually and electrically evoked responses must also be considered. For example, overlapping ON and OFF cells are frequently activated simultaneously in electrical stimulation, but this rarely happens with visual stimulation (but see (Turner and Rieke 2016)). If the parameters of the utility function $f(\cdot)$ are estimated from visually evoked responses, the utility function may be inaccurate for electrically evoked responses that are rarely if ever evoked by visual stimulation. A possible solution may be to use simpler models of the utility function (e.g. linear models), because they may be more robust to this problem than more complex and richer models (e.g. machine learning models) (Ben-David *et al* 2010).

Finally, multiple considerations suggest that the utility function $f(\cdot)$ may depend significantly on the location in the visual field (Wandell 1995, Rodieck 1998). Ideally, a retinal implant would be placed at the fovea, the region of the retina with the highest cell density and thus the highest visual acuity. However, many experiments have focused on the less dense peripheral retina, for technical reasons (Frechette *et al* 2005, Gauthier *et al* 2009, Turner and Rieke 2016) (but see (Wachtler *et al* 1996, Lee *et al* 2010, Sinha *et al* 2017)). In addition to the smaller receptive field sizes, differences in temporal integration in the cone photoreceptors and dominance of ON and OFF midget cell types have been reported in the fovea relative to the periphery (Dacey 2004, Sinha *et al* 2017). Further work is needed to fully characterize the unique visual response properties of the foveal region.

2.2. Identifying cell locations and types in a blind retina

A final challenge is to adapt our understanding of neural responses in healthy retinas to a blind retina. The most obvious approach is to identify the RGC locations and types in the target blind retina, and then

use the extensive body of retinal research to infer their original, healthy visual responses properties.

However, traditional approaches to cell type and location identification rely on light-evoked responses of RGCs and thus cannot be used in the retina of a blind person. Recent work shows that the cell locations and types in a given retina can often be identified from recordings of electrical activity without evaluating light responses (Richard *et al* 2015). Specifically, the electrophysiological image (EI) of a cell obtained with high-density recordings, which is measured by averaging the recorded voltage traces on each electrode across many spikes recorded from that cell, reveals structural and biophysical properties of the somatic, axonal and dendritic compartments of the cell (Litke *et al* 2004). First, the location of the soma and dendritic arbor provide an estimate of the receptive field location. Second, the sizes of the somatic and dendritic compartment and the axon conduction velocity can be particularly helpful in distinguishing parasol and midget cells (Li *et al* 2015), the most numerous classes of RGCs in the primate retina (Dacey 2004). Furthermore, additional differences in functional response properties such as firing rate (Sekirnjak *et al* 2011) and temporal structure of spike trains (Rhoades *et al* 2019) can be used to distinguish cell types, notably ON and OFF types within the midget and parasol cell classes (Sekirnjak *et al* 2011, Richard *et al* 2015). Together, these anatomical, biophysical and functional differences yield $\sim 85\%$ accuracy in distinguishing the five numerically dominant cell types in the primate retina (Richard *et al* 2015). While recent works have shown that the major RGC types in human retina are identical to those in macaques (Kling 2020), and previously developed methods for identifying these cell types from electrical activity are still applicable (Madugula 2020), further work is still needed to fully understand the translation to human retina (section 6).

After identifying the spatial properties of visual processing for each cell (the receptive field), the other parameters of light response must be also estimated to fully identify the utility function. Let us consider a simple utility function based on the linear-nonlinear-Poisson (LNP) encoding model, which predicts RGC light responses by passing the visual stimulus through a cascade consisting of a spatio-temporal linear filter followed by an instantaneous output nonlinearity and Poisson spike generation (Chichilnisky 2001). The temporal filter and output nonlinearity shapes are very similar within a cell type; hence, these parameters can be estimated for all the cells of a given type simultaneously (Chichilnisky and Kalmar 2002). To estimate these parameters in a blind retina, we can use cell-type specific parameters from previously recorded healthy retinas. However, there is substantial variation of these properties across different healthy retinas. Below, we describe an approach to address the problem of variability across retinas, not only for the

LNP encoding model described above, but also for a broader class of encoding models.

2.3. Characterizing diversity in neural responses

Most studies of neural processing in the retina focus on the features that are similar across retinas, often ignoring variability or treating it as random noise. However, visual response properties measured in many primate retinas exhibit clear structure in their variation (Shah 2019b). For simplicity, we describe our approach for studying this variability by modeling the utility function $f(\cdot)$ using a response encoding model $g(S)$ and a log-likelihood response similarity metric $\rho(\cdot)$, as described in equation (2). A multi-layered convolutional neural network (CNN) encoding model learned using data from 95 retinas failed to capture responses in all of the individual retinas simultaneously, because of inter-retina variability. To better capture this variability, a low-dimensional representation was learned with parameters to capture the individual properties of each retina in the collection, along with a single shared CNN with parameters common to all retinas (figure 2). In order to capture the responses in a particular retina, the different CNN layers were scaled using weights determined by the entries of a vector representing the location of the retina in the low-dimensional manifold (Shah 2019b). This manifold revealed a smooth variation in response properties such as the receptive field size, the dynamics of the temporal filter, the degree of output nonlinearity and the autocorrelation of responses, for two major cell types. The manifold also revealed surprising systematic differences in the visual response properties of RGCs in male versus female retinas. In addition to these variations, the manifold preserved the previously reported invariances across retinas, such as larger receptive field size and faster temporal integration of ON parasol cells compared to OFF parasol cells (Chichilnisky and Kalmar 2002).

The low-dimensional characterization of variability makes it possible to leverage hundreds of healthy retina recordings to efficiently identify the original visual responses in the retina of a blind person. The appropriate RGC encoding model, and hence a utility function $f(\cdot)$, can be identified by estimating an appropriate low-dimensional manifold location for the retina that the brain is expecting in a psychophysical discrimination task. At each step, the set of plausible manifold locations for characterizing visual behaviors is updated as follows: (i) sample a few of the plausible locations and generate corresponding retinal responses for a fixed visual stimulus, (ii) stimulate the artificial retina to reproduce these responses, (iii) query the subject about which response produces the most accurate sensation for a verbally described object, and (iv) update the set of plausible manifold locations. In simulations of this task, the set of plausible manifold locations converged to the location of the unknown retina in a small number of steps (Shah

2019b). Note, however, that the process could be substantially longer in a more realistic clinical application due to differences between the simulations using healthy primate retinas and the degenerated human retina (see section 6). In addition to the behavioral task, the manifold permits other approaches for estimating neural encoding in a previously unseen retina using any physiological measurements that may be available (figure 2).

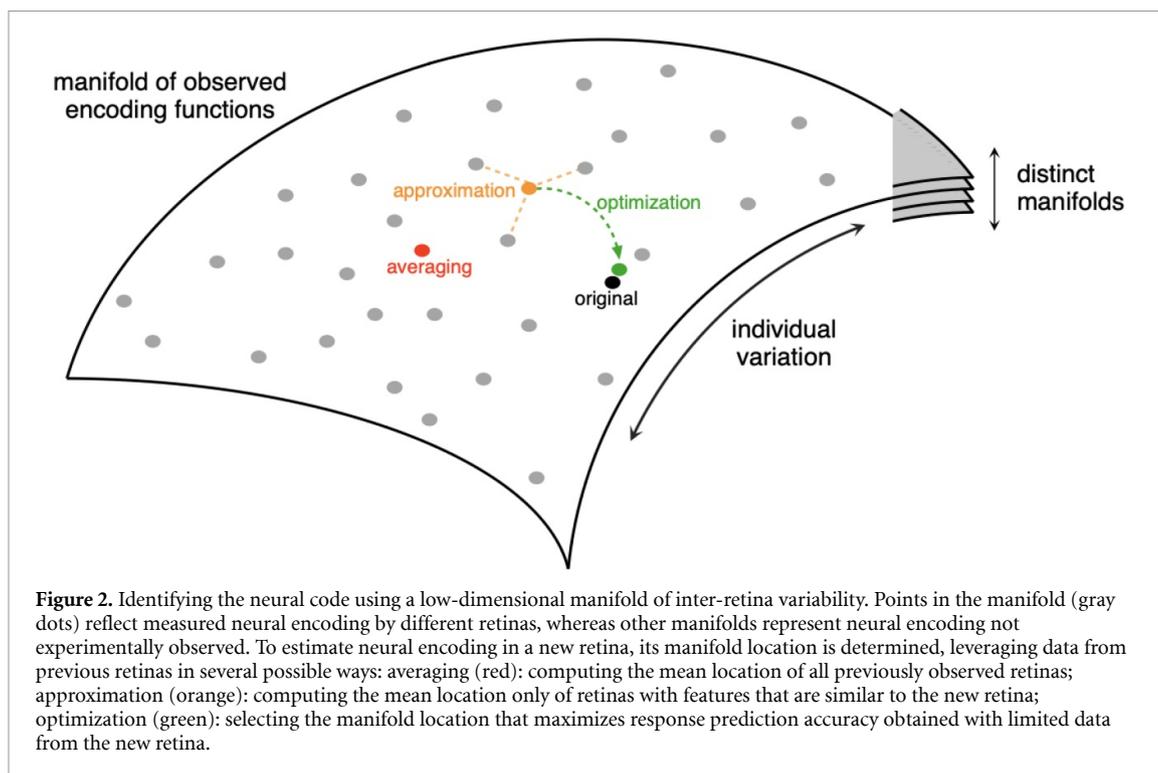
3. How does the retina respond to electrical stimulation?

The second major computational challenge is to measure how cells respond to electrical stimulation, $R \sim M(C)$ (figure 1). Ideally, the choice of electrical stimuli should be effective in reproducing the desired neural activity, the evoked responses should be accurately captured by a simple model, and the parameters of this model should be efficiently estimated using few measurements. To advance these goals, the problem of electrical response modeling can be broken down into several computational steps: choosing the electrical stimuli (section 3.1), identifying neural responses from recorded voltage traces after electrical stimulation (section 3.2, 3.3) and modeling the electrically evoked responses (section 3.4).

3.1. Choice of electrical stimuli

Ideally, the electrical stimuli used should be effective in reproducing the desired neural activity. Electrical stimulus consists of arbitrary current waveforms and amplitudes passed through an arbitrary combination of electrodes. Attempting to probe all possible stimuli produces a combinatorial explosion, thus, we start with a smaller, yet diverse collection of stimulation patterns. The simplest stimulation pattern is one in which a brief pulse of current (typically, charge-balanced, peak amplitude $\sim 1 \mu\text{A}$, duration $\sim 0.1 \text{ ms}$; (Sekirnjak *et al* 2007)) is passed through a single electrode. In our lab prototype, single-electrode stimulation using 512 electrodes with 40 distinct current amplitudes spans 40×512 current patterns, a small fraction of the total of 40^{512} possible patterns, thus avoiding the combinatorial explosion.

However, this simple starting point may not reveal the full ability of the device to replicate the neural code. Although an exhaustive exploration of more complex electrical stimuli is infeasible, the biophysics of electrical stimulation provides a guide to choosing a somewhat richer set of stimuli. One approach is to focus on selectivity, i.e. the ability to stimulate targeted cell(s) while avoiding others (Grosberg *et al* 2017, Madugula 2020). Several studies have shown that replacing the distant electrical ground in single-electrode stimulation with 'local return' electrodes can reduce the spread of the electric field, enhancing selectivity (Dommel *et al* 2005, Palanker *et al* 2005, Wong *et al* 2009, Abramian *et al* 2011, Joarder *et al*



2011, Cicione *et al* 2012, Habib *et al* 2013, Palanker 2014, Flores *et al* 2016, Sinclair *et al* 2016, Spencer *et al* 2016), including in recent work performed at cellular resolution (Fan *et al* 2019). Also, more general ‘current steering’ methods aim to improve selectivity further by passing currents through multiple nearby electrodes in a way that systematically shifts the spatial focus of electrical activation (Townshend and White 1987, Sweeney *et al* 1990, Bonham and Litvak 2008, Butson and McIntyre 2008, Martens *et al* 2011, Habib *et al* 2013, Matteucci *et al* 2013, Spencer *et al* 2018). Similarly, other methods modify the temporal properties of activation by trading off current amplitude with pulse duration (Boinagrov *et al* 2010, 2014, Rattay *et al* 2012, Lee *et al* 2013, Weitz *et al* 2015). Note that although most of these electrical stimuli have been developed for improved selectivity, it is possible that aspects other than selectivity are more important for reproducing the desired visual sensations (see section 4).

3.2. Spike sorting in the presence of electrical artifact

Determining how effectively the electrical stimulus activates each cell is accomplished by passing current, recording the resulting voltage traces on all the electrodes, and identifying any elicited spikes. Unfortunately, the recorded voltage traces are typically corrupted by electrical stimulation artifacts, the properties of which are a complex function of the hardware, tissue and the electrical stimulus itself. The goal of spike sorting in this setting is to estimate and subtract stimulation artifacts to obtain the true physiological signal, and then identify in this

signal the waveforms of spikes produced by cells that were previously identified in the artifact-free spontaneous activity. Typically, manual spike sorting to identify the responses to a particular electrical stimulus over a range of amplitudes requires several minutes for a trained human. For thousands of electrodes and multiple cells that are potentially activated by each electrode, manual spike sorting each electrode-cell pair is extremely time-consuming. Hence, automation of this step is important, but it is challenging (Hashimoto *et al* 2002, Wagenaar and Potter 2002, Heffer and Fallon 2008, Erez *et al* 2010). In one automated approach applied to single-electrode stimulation, a prior on artifact shape to enforce smooth changes across successive current amplitudes, and a fast binary pursuit spike matching approach, yielded >90% accuracy in estimating the activation probability compared to manual analysis (Mena *et al* 2017). Modification of this spike sorting algorithm for more complex stimulation patterns would require a better understanding of the artifact shape and how it varies for a wide range of electrical stimuli. An ideal alternative would be to find electrical stimuli that are effective at eliciting desired activity but also have much smaller stimulation artifacts, simplifying spike sorting (Brown *et al* 2008, Wichmann and Devergnas 2011, Hottowy *et al* 2012).

3.3. Identifying and avoiding axonal activation of distant cells

The approach of electrically stimulating each cell to match its healthy visually evoked responses is not applicable when the location of the cell body, and

thus the location of its original visual receptive field, cannot be determined. This happens with electrical stimuli that activate the axon of a RGC with a cell body that is away from the recording array and which thus cannot be located. Indeed, in existing clinical implants (Humayun *et al* 2012), simultaneous activation of many cells via their axons results in large, arc-shaped percepts determined by the distant locations of their cell bodies (Nanduri 2011). Detecting and avoiding this problem can be accomplished by exploiting the unique spatio-temporal footprint of axonal activation on a high-density electrode array: a voltage waveform originating at the stimulation site and propagating bi-directionally along the axon's trajectory (Grosberg *et al* 2017, Tandon *et al* 2017). Note that this technique is particularly effective in the retina and other systems in which the prevalent direction of axons is known.

3.4. Modeling the response to electrical stimulation

To fully exploit the capabilities of a high-resolution artificial retina, it would be helpful to have a simple description of the relationship between electrical stimuli and spiking activity, in order to predict RGC responses to arbitrary stimulation patterns. For single-electrode stimulation, a sigmoidal relationship between current amplitude and the activation probability of a given cell has been effective (Sekirnjak *et al* 2008, Fried *et al* 2009, Jepson *et al* 2013). However, both the threshold and sensitivity of this relationship can vary greatly at different locations near the cell body, axon, and dendrites (figure 3), and for large current values, inactivation of cells at high current levels may require a non-monotonic functional form (Meng *et al* 2018). As with the choice of electrical stimuli (see section 3.1), modeling of electrical responses can benefit from biophysical insights, such as the different electrical properties of the axon and soma (Rattay 1986, 1989, Rattay *et al* 2012). However, electrical responses also depend on properties that are not easily measured, such as the spatial relationship between the cell and electrode, and the anisotropic extracellular environment consisting of axon fibers, other cells, and the inner limiting membrane. Hence, the benefit of exact biophysical models for understanding extracellular stimulation is less obvious than it is for intracellular stimulation.

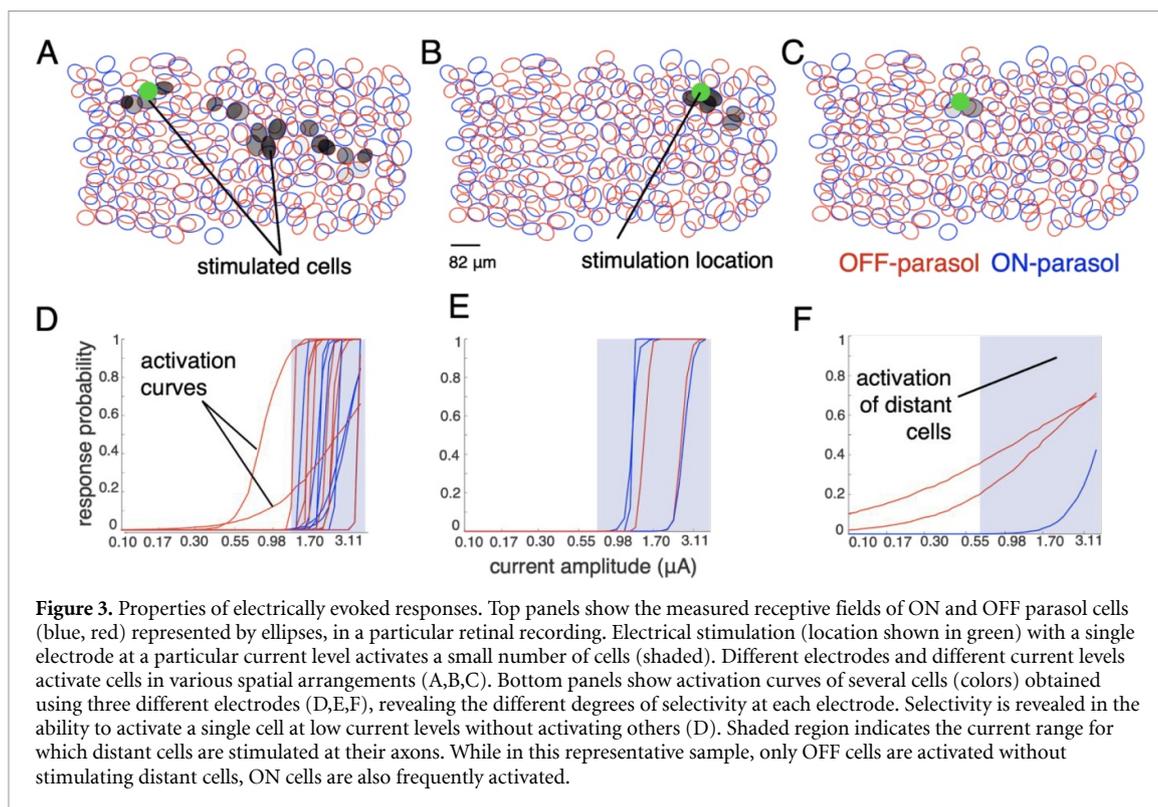
A possible alternative is to derive the mathematical form of the relationship between current and cellular activation using biophysical models, and then fit the unknown model parameters using data obtained by stimulating and recording. This approach has been useful for comparing somatic and axonal stimulation (Fohlmeister and Miller 1997, Rattay 1999) and for characterizing the efficacy of stimulation with current passed through two nearby electrodes (Jepson *et al* 2014b). In the latter study, biophysical evidence suggests that the RGC response probability can be modeled as a combination of sigmoids, each over a

different linear combination of stimulation currents (Esler *et al* 2018). Each sigmoid is hypothesized to be associated with a distinct spike initiation site on the cell, depending on which electrode dominates activation. Each spike initiation site is associated with a different linear trade-off in the current passed through the two electrodes to reach activation threshold, resulting in a piecewise linear iso-response curve (Jepson *et al* 2014b).

In the future, electrical response models could build upon methods that have been used to estimate visual response models. For example, electrical response models for bi-electrode stimulation can be fitted to experimental data using techniques developed for estimating visual receptive field 'subunits' (Kaardal *et al* 2013, McFarland *et al* 2013, Freeman *et al* 2015, Maheswaranathan *et al* 2018, Shah *et al* 2020). These subunits affect neural responses through a nonlinear combination of linear summations of inputs, broadly similar to the effect of multiple electrical activation sites, though the underlying mechanism is different (synaptic rectification due nonlinear biophysics of transmitter release). In addition to the model architecture, tailored stimuli for the efficient estimation of model parameters can also build on methods developed for visual response characterization. Recently, the method used for identifying an LNP visual response model using white noise stimuli (Chichilnisky 2001) has been adapted for electrical stimuli (Halupka *et al* 2017, Ho *et al* 2018, Rathbun *et al* 2018).

3.5. Efficient calibration

In all of the above approaches, efficiency is critical. For example, in the laboratory prototype experiments, measurement of only the responses to single-electrode stimulation (512 electrodes X 40 current amplitudes) requires ~ 1.5 h of experimentation, and calibrating the evoked responses to a large fraction of the distinct possible multi-electrode stimulation patterns is not feasible. One approach to increasing efficiency is to adaptively select stimulation patterns based on previous measurements. In the case of single-electrode stimulation, it would be natural to focus on intermediate current levels that lead to activation probability near 0.5, to rapidly constrain an estimate of the sigmoidal activation curve that relates current to spike probability. However, doing this in closed-loop requires computationally expensive steps of spike sorting and response modeling with each measurement. An alternative approach with fewer computational requirements is to use electrical stimulation properties measured in previous recordings as priors. For spike sorting, the similarity of recorded spikes and artifacts across experiments can be exploited to reduce the dimensionality of the problem (Shah *et al* 2019a). For response modeling, the inverse relationship between recorded spike amplitude and activation threshold can be



used to estimate activation curves with fewer trials (Madugula *et al* 2019). Importantly, using these priors reduces the required number of measurements by an amount similar to the reduction produced by adaptive stimulation, without requiring closed-loop computation (Shah *et al* 2019a). Additional work will be needed to understand the benefits of these priors in the presence of possible implant instability over time or greater diversity of responses across cells in a blind person.

4. How do we optimally replicate RGC firing in a blind retina?

A third major computational problem for an advanced artificial retina is using the visual and electrical response properties determined above to optimize electrical stimulation, for a given target visual stimulus. Ideally, we aim to reproduce the desired neural activity cell-by-cell and spike-by-spike. However, even the most advanced neural interfaces are likely to have imperfect selectivity, and it is not *a priori* obvious how to measure and optimize performance when the desired neural responses cannot be replicated exactly. This is because different kinds of stimulation errors would be expected to influence visual perception in different ways. For example, due to the spatial correlations in natural images, it may often be acceptable to simultaneously activate two nearby cells of the same type with little loss of resolution, but activating cells of different types (e.g. ON and OFF cells) may be more problematic. Hence, it is desirable to identify a utility function $f(S, R)$ that

measures device performance in a way that is related to the accuracy of the elicited visual perception. In the above example, the utility function would typically not greatly penalize the simultaneous activation of the two nearby cells of the same type, but would penalize activation of different cell types more strongly.

Given an appropriate utility function, the next challenge is optimizing it in real time, under the constraints of the neural interface. This problem can be solved using standard methods if there exists an electrical response model M that correctly predicts the neural responses evoked by arbitrary electrical stimuli. However, as described in section 3, M is difficult to estimate from measurements with a limited collection of electrical stimuli.

One approach is to exploit the relatively slow integration time of the visual system by rapidly presenting a diverse sequence of electrical stimuli from a limited, calibrated dictionary, so that the elicited responses are effectively summed in downstream neural circuits to produce accurate visual perception. Note that the calibrated dictionary allows both axonal and somatic activation, but avoids high current levels that produce off-array axonal activation, because in these cases the receptive location, and hence the desired visual response, cannot be identified. The temporal dithering approach exploits the fact that electrical stimulation hardware is fast (~ 10 kHz) and evoked spike latencies in response to direct electrical stimulation are precise (~ 0.1 ms), while synaptic integration of neural inputs downstream is comparatively slow (e.g. ~ 10 ms) (see figure 4) (Sekirnjak *et al* 2008, Jepson *et al* 2013,

Kandel 2013, Shah *et al* 2019a). The underlying assumptions are that the evoked RGC responses are generated independently over time during the rapid stimulation sequence, and that the resulting visual perception depends only on the integrated neural activity downstream. With this assumption, the problem in equation (1) reduces to finding a sequence of dictionary elements (C_1, \dots, C_n ; n is sequence length), such that the summation of corresponding probabilistically generated responses ($\sum_{i=1}^{i=n} R_i$) maximizes the expected utility function f with the target visual stimulus S :

$$C_1, \dots, C_n = \underset{\text{argmax}}{\mathbb{E}_{R_i \sim M(C_i)}} f\left(S, \sum_{i=1}^{i=n} R_i\right) \quad (5)$$

With this framework, optimization of the device comes down to selecting the appropriate stimulation sequence in real time. Perhaps the simplest approach is a *greedy* one, in which an electrical stimulus is chosen at each time step such that it maximizes the increment to the utility function. We refer to this approach as greedy temporal dithering (GTD).

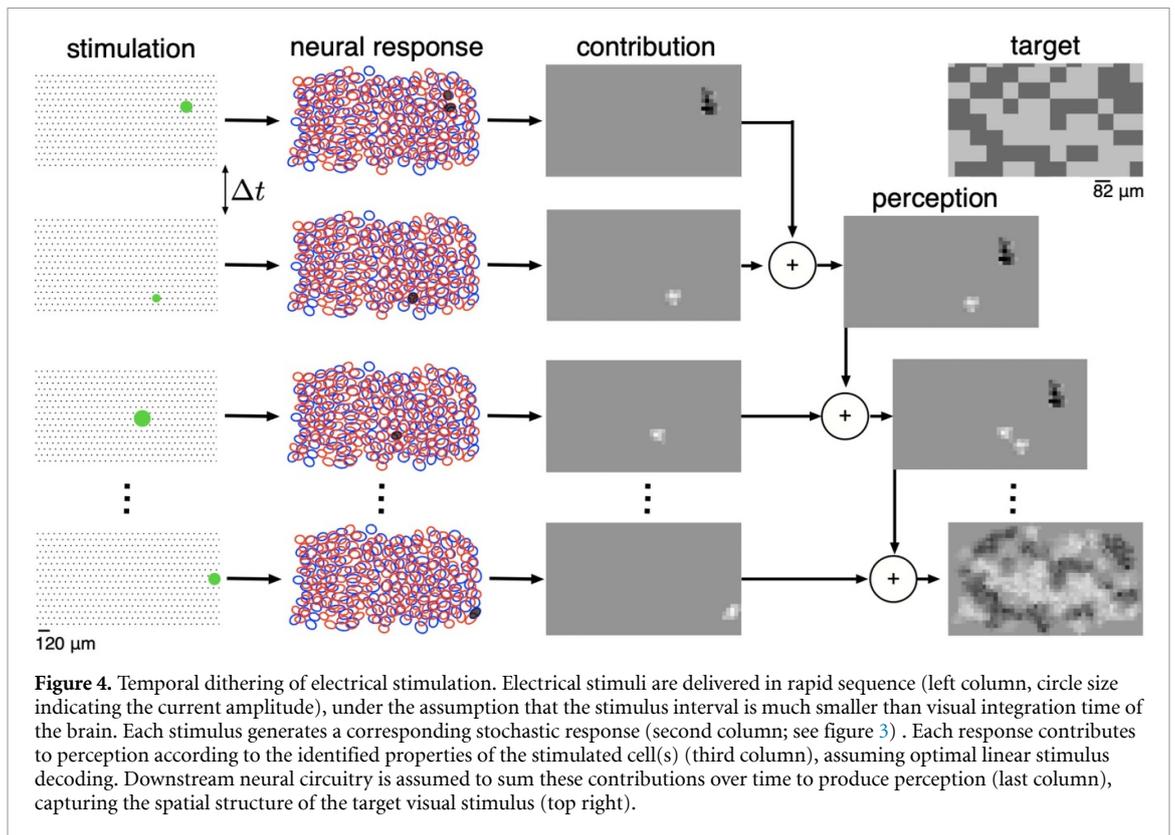
The visual perception produced by the GTD approach was analyzed using the experimental lab prototype, with data from ON and OFF parasol cells in isolated (healthy) primate retina (figure 4) (Shah *et al* 2019a). Visually evoked responses were used to estimate the parameters of the utility function $f(\cdot)$, assuming as a cost the mean-squared error between the target visual stimulus and the predicted perception obtained by linearly decoding the RGC responses evoked by electrical stimulation. A coarse white noise stimulus, which has some of the spatial correlations in natural images due to large pixel size, was used to estimate the optimal linear decoding filters. Electrically evoked responses were measured using a dictionary of single-electrode stimulation patterns: each dictionary element corresponded to current passed through one of the 512 electrodes at one of 40 amplitudes. For each dictionary element, the response probabilities for every cell were estimated by applying automated spike sorting (Mena *et al* 2017) to recordings of multiple presentations of the electrical stimulus. Then, for a target visual image, a sequence of dictionary elements was selected in rapid succession, to greedily maximize the increment to the utility function. Simulated RGC responses to this sequence were generated using the calibrated (measured) activation probabilities associated with each electrical stimulus. The resulting visual perception was estimated by linearly decoding the image from these simulated RGC responses using the spike-triggered average of each cell as the decoding filter. Across multiple target images and sampled responses, the GTD algorithm showed nearly optimal performance compared to the best possible temporal dithering sequence (Shah *et al* 2019a, Madugula 2020). This result implies that for the development of an implanted device, more

accurate visual restoration performance does not rely on better algorithms, but instead requires a higher resolution interface with the retina to produce a richer stimulation dictionary. Further work is needed to experimentally validate the main assumptions behind the GTD approach, notably the independence and additivity of responses elicited by successive electrical stimuli.

Surprisingly, in this test the GTD algorithm tended to select electrodes that activated multiple cells simultaneously, rather than one cell selectively (Shah *et al* 2019a). This observation is in contrast to previous efforts to optimize the selectivity of activation, i.e. activating one or a few target cells while not activating non-target cells (see section 3 above) (Fried *et al* 2006, Wong *et al* 2009, Freeman *et al* 2010, Jepson *et al* 2013, 2014b, Lee and Im 2019). Note that the preference for less selective electrodes was also observed using an alternative algorithm that provides a lower bound for the optimal algorithm; hence, the preference of non-selective stimulation is not merely a consequence of the greedy selection procedure. A possible interpretation is that when perfect selectivity is not possible, it is often beneficial to exploit the simultaneous activation of multiple nearby cells. For example, this could be helpful when the incoming image has a spatial structure which is well-matched to the collection of cells activated by a particular electrode. The idea that selectivity is not necessarily the most important goal could be further examined by applying the GTD approach using more selective electrical stimuli, e.g. those with local returns (see section 3.1 above), and testing the impact on decoding. Because the GTD framework and lab prototype permit this kind of simulation of visual perception, the approach can also be used to study the robustness of electrically evoked responses to phenomena such as array movement, cell death and changes in stimulation efficacy over time.

Applying the GTD framework to data from the lab prototype also provides novel insights into the design of an implantable device. For example, restricting stimulation to the 50% of the most frequently used electrodes resulted in a negligible decrease in overall performance. This observation suggests an adaptive design, in which a high-density electrode array is implanted, and half of the electrodes are subsequently turned off based on the measured electrical response properties, resulting in reduced power consumption and heat dissipation.

While the GTD framework is described above for a static visual target, the approach can be extended to natural vision. In natural viewing, the visual scene is scanned by eye movements that position the fovea at many different locations to obtain high-resolution visual information. Similarly, because the implant is small compared to the entire visual scene, it is expected that the implant user will scan the visual scene with eye movements (Sabbah *et al* 2014). To simulate



this process, the GTD framework can be applied in sequence at every saccade location with the corresponding region of visual scene as the target, and the overall perception of the scene can be simulated by summing the reconstructed image at these locations over space and time.

5. System design for implementing the computational steps

To translate these computational insights into greater acuity of artificial vision, the algorithms must be implemented within hardware constraints such as power, area and communication bandwidth and latency. Figure 5 presents a system-level implementation consisting of three components: (i) an external processor with a video camera and eye position sensor, (ii) a data and power relay, mounted on the outside of the eye, and (iii) an electrode array interface implanted on the retina. Compared to the external processor, the implanted array has severely constrained area and power limits, for surgical and safety reasons ($<1 \text{ mW mm}^{-2}$) (Mathieson *et al* 2012), as well as limited bandwidth and latency for wireless data and power transmission, because of the variable efficiency of the wireless link as the eye scans the visual field. The relay mitigates these limitations by providing power for intermediate computations and improving the efficiency of wireless transmission by being close to the processor and not moving relative to the implant. For visual and electrical response calibration, the recorded data are transmitted from

the implant via the relay to the processor. Conversely, during runtime function, the target visual stimulus is captured by the external camera, and information about the stimulation sequence is sent from the processor via the relay to the interface.

A distributed implementation of the algorithms described earlier across different hardware components may enable continuous operation of the artificial retina within the constraints of power, processing, memory, and communication. For example, assuming a moderate communication bandwidth between the processor and the relay, and a lower bandwidth between the relay and implant, a possible distributed implementation of the GTD algorithm could be: (i) select a small subset of the electrical stimulation dictionary on the external processor, based on the target image and the measured eye position immediately after a saccade; (ii) send this reduced dictionary to the relay; (iii) use the reduced dictionary to compute the stimulation sequence on the relay; and (iv) send the stimulation sequence in real time to the interface. Further improvement is possible by pre-computing the dictionary for various saccade locations.

If implementation of the proposed algorithms is difficult within the constraints of existing hardware, co-design of hardware and algorithms may permit a smoother trade-off between efficiency and accuracy. A potential example of hardware-algorithm co-design is based on the properties of eye movements that are crucial for normal visual perception. Recent studies have shown that the brain uses

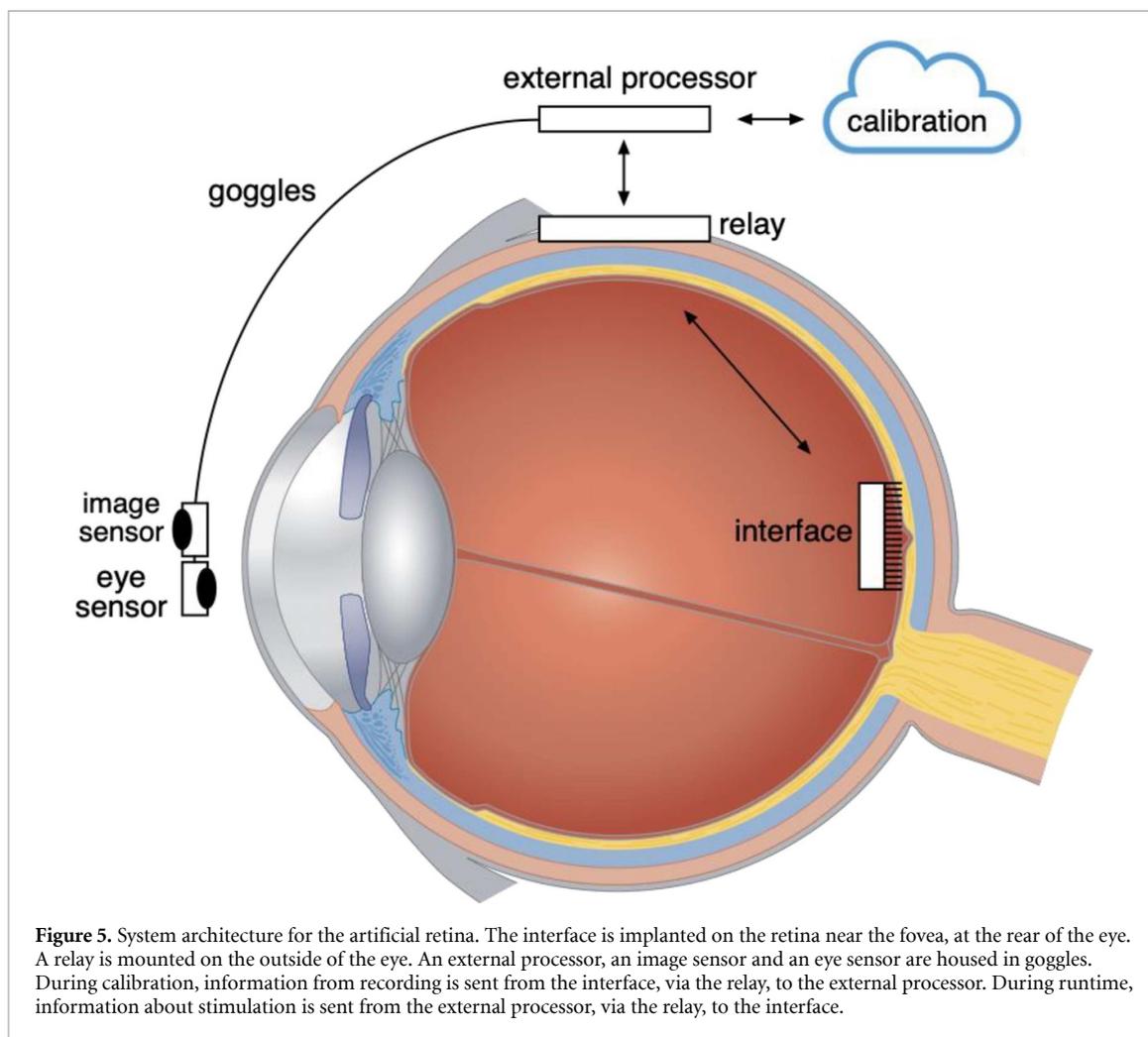


Figure 5. System architecture for the artificial retina. The interface is implanted on the retina near the fovea, at the rear of the eye. A relay is mounted on the outside of the eye. An external processor, an image sensor and an eye sensor are housed in goggles. During calibration, information from recording is sent from the interface, via the relay, to the external processor. During runtime, information about stimulation is sent from the external processor, via the relay, to the interface.

the statistics of microsaccadic eye movements, not their exact trajectory, to decode visual stimuli from RGC responses (Ratnam *et al* 2017). This suggests the possibility of relaxed hardware requirements for the precision of measured eye movements, because adding a simulated (rather than measured) microsaccade trajectory to the coarsely measured saccade trajectory may suffice for high-resolution artificial vision.

Another example of hardware-algorithm co-design is the development of compression for electrical recordings at the neural interface, to reduce data processing and communication bandwidth. Naive front-end lossy compression approaches could backfire by compromising subsequent computational steps, such as spike sorting and cell-type classification. Instead, co-designing the hardware and algorithm using data from the lab prototype allows a balance between compression and performance of the downstream computational steps. Recently it has been shown that exploiting the sparsity of spike data by projecting analog voltage traces non-uniquely across the rows and columns of the interface electrodes can produce $\sim 40X$ compression with minimal loss of relevant spike data (Muratore *et al* 2019). Similarly, spike sorting algorithms for electrical

stimulation can benefit from hardware innovations resulting in reduced artifacts or better priors on the artifact waveform. Thus, system and hardware design may be improved using insights obtained using the lab prototype.

6. Experimental limitations and caveats

The work described here builds on experimental data obtained from healthy isolated non-human primate retinas, using a laboratory prototype system for high-density recording and stimulation. Although this system provides major insights, it differs in important ways from the clinical goal: an artificial retina implanted in the eye of blind human subjects. Hence, future work must focus on computational approaches that are robust to these differences.

First, this work focused on macaque monkey retinas, because the gross structure, collection of cell types, and visual capabilities and behaviors of macaques are closer to those of humans than any other animal model. Indeed, recent physiological work on human RGCs (Cowan 2019, Kling 2020, Soto 2020) shows that the functional properties of the corresponding cell types in the two species are as similar as their morphology suggests (Dacey 1993)

(but see Reinhard and Münch 2019). Furthermore, the focal electrical stimulation properties are also very similar (Madugula 2020). In the present context, the computational goal would be to understand the normal function of a given human retina as a sample on the manifold obtained using many primate retinas (section 2.3), so that the similarities between human and macaque retinas can be exploited for an effective clinical implant 2.3.

Second, during retinal degeneration, significant changes in network (Jones *et al* 2016) and intrinsic RGC properties could affect the artificial visual signal transmitted to the brain (Spencer *et al* 2018). Compared to stimulation methods that interface to the upstream circuitry, the direct epiretinal stimulation approach could be more robust to these changes, but challenges remain. Previous studies have reported oscillatory activity (Goo *et al* 2015) and changes in spontaneous activity (Trenholm and Awatramani 2015) in rodent models of retinal degeneration. Furthermore, in extreme cases of retinal degeneration, the excitable RGCs are found in patches, with only a small fraction of cells remaining in extreme cases (Ren *et al* 2018, García-Ayuso and Di Pierdomenico 2019). The impact of these changes on the present methods is uncertain. Adding evoked spikes to spontaneous spikes could produce a visual signal that is contaminated by noise, however, it is possible that spontaneous firing will return to normal after extended ongoing electrical stimulation, and future approaches may make it possible to suppress firing with electrical stimulation. Degenerated patches of RGCs could create permanent islands of missing visual function, however, it is possible that eye movements will compensate by directing the image to intact locations. Resolving these concerns will require tests in primate models of degeneration.

Third, the methods used for obtaining high quality recordings using isolated retinas may not apply directly to the *in vivo* setting. For example, novel methods must be developed to establish close electrical contact between the implant and the retina. Additionally, if the implant or the retina are not stable over long durations (which cannot be tested with isolated retina), frequent re-calibration of electrical and visual response properties may be required.

Finally, results obtained in the peripheral retina (where most recordings are performed) should be interpreted with caution, because calibration of electrical responses will likely depend strongly on the region of the retina that is targeted and the electrical contact between the device and retina (see section 2). Although the central retina is an attractive clinical target because the higher density of cells provides higher resolution natural vision, high cell density could also make it harder to selectively stimulate a targeted cell electrically while avoiding other nearby cells. These considerations raise a key question for device placement: does stimulation with low selectivity in the

central retina produce more or less faithful vision restoration than stimulation with high selectivity in a more peripheral location? The raphe region of the retina ($\sim 2\text{--}5$ mm from the fovea on the horizontal visual meridian; roughly $8\text{--}20^\circ$ eccentricity), presents a possible compromise: in the raphe, cell density is relatively high, but RGCs do not form multiple layers as they do in the fovea, leading to easier electrical access using a flat electrode array. Furthermore, because axons avoid the raphe region on their way from the peripheral retina to the optic disk, activation of distant cells via their axons is less of a problem (Grosberg *et al* 2017). Investigation of the electrical and visual response properties of the raphe and surrounding regions is an important topic for future investigation (Gogliettino *et al* 2019).

7. Broader implications and future work

Once a high-resolution, bi-directional artificial retina is built, the computational steps described here may guide new research into the mechanisms of visual processing. Examples include comparing the artificial vision produced by different RGC types and probing the perception resulting from the interaction between natural and artificial vision (Arens-Arad *et al* 2020). Moreover, the presented framework also raises new questions about the relationship between advances in the basic science knowledge of visual processing and the engineering of a neural interface. What is the simplest visual utility function (f), that can be implemented efficiently in hardware, for a given electrical response model (M)? Conversely, what choices of electrical stimuli yield high performance for a variety of visual utility functions (f) and hardware implementation constraints?

The computational problems and methods introduced here are specific and concrete, in large part because the retina is a relatively well-understood neural circuit with well-defined function in vision. Related concepts may be applicable to other neural circuits, such as those mediating auditory, visual, or somatosensory capacities, and to other recording and stimulation technologies, such as optical, ultrasound or magnetic approaches. Adapting these ideas to different neural systems will depend on identifying the corresponding utility function (f), either as an encoding model, a decoding model or a joint embedding of stimuli and responses. Similarly, adapting to different recording and stimulation modalities may be possible by first measuring the corresponding response model (M) and compensating for possible non-selectivity and the high-dimensional space of neural stimulation patterns using temporal dithering.

Future work on the artificial retina and other neural interfaces may require revisiting some of the assumptions and the technical choices made in this work. First, we have proposed stimulation choices based on pre-calibration obtained with

recording. An ideal high-performance closed-loop system (Krook-Magnuson *et al* 2015) could further compensate for the stochasticity of electrical stimulation by passing current, recording the resulting voltages, identifying the activated cells, and modifying the subsequent electrical stimulus, all in real time. The simpler approach presented here involves choosing electrical stimuli based on improvement in the *expected* utility function, averaged over possible patterns of evoked neural responses, based on a calibration performed in advance. While this approach is less computationally intensive and easier to implement in hardware, it is unsuitable if the expected utility function diverges significantly from evoked responses on individual trials. In the artificial retina application, because each RGC is activated independently of other RGCs and the ‘memory’ of electrical stimulation is limited (sub-millisecond), the utility function on individual trials is close to the expected utility function. However, the use of the expected utility function should be revisited for application in other neural circuits, where network-mediated recurrence could lead to increased inter-trial variability and interactions between stimulated neurons.

Second, we have assumed that the utility function is *stationary*, i.e. visual response properties do not change over time. However, some degree of plasticity in visual response properties has been previously observed, for example, after retinal detachment and reattachment. Future work may need to address plasticity by adjusting to changes in the utility function. Furthermore, it may be possible to gradually drive plasticity with electrical stimulation, such that downstream processing adjusts to the properties of the implant. In other neural circuits, such as the cortex, plasticity may be more prominent and thus these issues may be more central. The simpler situation in the retina thus provides an important stepping stone in the design of bi-directional and high-resolution interfaces to other areas of the brain.

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ORCID iD

Nishal P Shah  <https://orcid.org/0000-0002-1275-0381>

References

- Abramian M, Lovell N H, Morley J W, Suaning G J and Dokos S 2011 Activation of retinal ganglion cells following epiretinal electrical stimulation with hexagonally arranged bipolar electrodes *J. Neural Eng.* **8** 035004
- Anumanchipalli G K, Chartier J and Chang E F 2019 Speech synthesis from neural decoding of spoken sentences *Nature* **568** 493–8
- Arens-Arad T, Farah N, Lender R, Moshkovitz A, Flores T, Palanker D and Mandel Y 2020 Cortical interactions between prosthetic and natural vision *Curr. Biol.* **30** 176–182.e2
- Ayton L N *et al* 2014 First-in-human trial of a novel suprachoroidal retinal prosthesis *PLoS One* **9** e115239
- Bacher D, Jarosiewicz B, Masse N Y, Stavisky S D, Simeral J D, Newell K, Oakley E M, Cash S S, Friehs G and Hochberg L R 2015 Neural point-and-click communication by a person with incomplete locked-in syndrome *Neurorehabil. Neural Repair* **29** 462–71
- Batty E *et al* 2016 Multilayer Recurrent Network Models of Primate Retinal Ganglion Cell Responses (Available at <https://openreview.net/pdf?id=HkEI22jeg>) (Accessed: 28 March 2020)
- Ben-David S, Blitzer J, Crammer K, Kulesza A, Pereira F and Vaughan J W 2010 A theory of learning from different domains *Mach. Learn.* **79** 151–75
- Beyeler M *et al* 2019 A model of ganglion axon pathways accounts for percepts elicited by retinal implants *Sci. Rep.* **9** 9199
- Boinagrov D, Loudin J and Palanker D 2010 Strength–duration relationship for extracellular neural stimulation: numerical and analytical models *J. Neurophysiol.* **104** 2236–48
- Boinagrov D, Pangratz-Fuehrer S, Goetz G and Palanker D 2014 Selectivity of direct and network-mediated stimulation of the retinal ganglion cells with epi-, sub- and intraretinal electrodes *J. Neural Eng.* **11** 026008
- Bonham B H and Litvak L M 2008 Current focusing and steering: modeling, physiology, and psychophysics *Hearing Res.* **242** 141–53
- Brackbill N *et al* 2017 Reconstruction of natural images from responses of primate retinal ganglion cells *European Retina Meeting* (accepted) (<https://doi.org/10.1101/2020.05.04.077693>)
- Brown E A *et al* 2008 Stimulus-artifact elimination in a multi-electrode system *IEEE Trans. Biomed. Circuits Syst.* **2** 10–21
- Butson C R and McIntyre C C 2008 Current steering to control the volume of tissue activated during deep brain stimulation *Brain Stimul.* **1** 7–15
- Carandini M and Heeger D J 2011 Normalization as a canonical neural computation *Nat. Rev. Neurosci.* **13** 51–62
- Chander D and Chichilnisky E J 2001 Adaptation to temporal contrast in primate and salamander retina *J. Neurosci.* **21** 9904–16
- Chichilnisky E J 2001 A simple white noise analysis of neuronal light responses *Network* **12** 199–213
- Chichilnisky E J and Kalmar R S 2002 Functional asymmetries in ON and OFF ganglion cells of primate retina *J. Neurosci.* **22** 2737–47
- Cicione R, Shivdasani M N, Fallon J B, Luu C D, Allen P J, Rathbone G D, Shepherd R K and Williams C E 2012 Visual cortex responses to suprachoroidal electrical stimulation of the retina: effects of electrode return configuration *J. Neural Eng.* **9** 036009
- Cowan C S *et al* 2019 Cell types of the human retina and its organoids at single-cell resolution: developmental

- convergence, transcriptomic identity, and disease map *Cell* **182** 1623–40
- Dacey D M 1993 The mosaic of midget ganglion cells in the human retina *J. Neurosci.* **13** 5334–55
- Dacey D 2004 20 origins of perception: retinal ganglion cell diversity and the creation of parallel visual pathways *The Cognitive Neurosciences III* M S Gazzaniga ed (Cambridge, MA: MIT Press) p 281
- Demb J B *et al* 2001 Bipolar cells contribute to nonlinear spatial summation in the brisk-transient (Y) ganglion cell in mammalian retina *J. Neurosci.* **7447–54**
- Demb J B, Haarsma L, Freed M A and Sterling P 1999 Functional circuitry of the retinal ganglion cell's nonlinear receptive field *J. Neurosci.* **19** 9756–67
- Dommel N *et al* 2005 *In-vitro* testing of simultaneous charge injection and recovery in a retinal neuroprosthesis. 2005 *IEEE Eng. Med. Biol. 27th Annual Conf.* pp 7612–5
- Dowling J E 1987 *The Retina: An Approachable Part of the Brain* (Cambridge, MA: Harvard University Press)
- Dragas J *et al* 2017 *In vitro* multi-functional microelectrode array featuring 59 760 electrodes, 2048 electrophysiology channels, stimulation, impedance measurement, and neurotransmitter detection channels *IEEE J. Solid State Circuits* **1576–90**
- Dubbs A J, Seiler B A and Magnasco M O 2010 A fast L_2 spike alignment metric *Neural Comput.* **2785–808**
- Erez Y, Tischler H, Moran A and Bar-Gad I 2010 Generalized framework for stimulus artifact removal *J. Neurosci. Methods* **191** 45–59
- Esler T B, Maturana M I, Kerr R R, Grayden D B, Burkitt A N and Meffin H 2018 Biophysical basis of the linear electrical receptive fields of retinal ganglion cells *J. Neural. Eng.* **15** 055001
- Fan V H, Grosberg L E, Madugula S S, Hottoway P, Dabrowski W, Sher A, Litke A M and Chichilnisky E J 2019 Epiretinal stimulation with local returns enhances selectivity at cellular resolution *J. Neural. Eng.* **16** 025001
- Flores T, Goetz G, Lei X and Palanker D 2016 Optimization of return electrodes in neurostimulating arrays *J. Neural. Eng.* **13** 036010
- Fohlmeister J F and Miller R F 1997 Mechanisms by which cell geometry controls repetitive impulse firing in retinal ganglion cells *J. Neurophysiol.* **78** 1948–64
- Frechette E S, Sher A, Grivich M I, Petrusca D, Litke A M and Chichilnisky E J 2005 Fidelity of the ensemble code for visual motion in primate retina *J. Neurophysiol.* **94** 119–35
- Freeman D K, Eddington D K, Rizzo J F and Fried S I 2010 Selective activation of neuronal targets with sinusoidal electric stimulation *J. Neurophysiol.* **104** 2778–91
- Freeman J *et al* 2015 Mapping nonlinear receptive field structure in primate retina at single cone resolution *eLife* **4** e05241
- Fried S I, Hsueh H A and Werblin F S 2006 A method for generating precise temporal patterns of retinal spiking using prosthetic stimulation *J. Neurophysiol.* **95** 970–8
- Fried S I, Lasker A C W, Desai N J, Eddington D K and Rizzo J F 2009 Axonal sodium-channel bands shape the response to electric stimulation in retinal ganglion cells *J. Neurophysiol.* **101** 1972–87
- Ganmor E, Segev R and Schneidman E 2015 A thesaurus for a neural population code *eLife* **4** e06134
- García-Ayuso D and Di Pierdomenico J 2019 Retinal ganglion cell death as a late remodeling effect of photoreceptor degeneration *Int. J. Mdpi* **20** 4649
- Gauthier J L, Field G D, Sher A, Greschner M, Shlens J, Litke A M and Chichilnisky E J 2009 Receptive fields in primate retina are coordinated to sample visual space more uniformly *PLoS Biol.* **7** e1000063
- Goetz G A and Palanker D V 2016 Electronic approaches to restoration of sight *Rep. Prog. Phys.* **79** 096701
- Gogliettino A R *et al* 2019 Electrical recording and stimulation of RGCs in the macaque raphe at cellular resolution in *The Eye and the Chip (TEATC)*
- Golden J R, Erickson-Davis C, Cottaris N P, Parthasarathy N, Rieke F, Brainard D H, Wandell B A and Chichilnisky E J 2019 Simulation of visual perception and learning with a retinal prosthesis *J. Neural. Eng.* **16** 025003
- Gollisch T and Meister M 2010 Eye smarter than scientists believed: neural computations in circuits of the retina *Neuron* **65** 150–64
- Goo Y S *et al* 2015 Spontaneous oscillatory rhythms in the degenerating mouse retina modulate retinal ganglion cell responses to electrical stimulation *Frontiers Cell Neurosci.* **9** 512
- Greschner M, Shlens J, Bakolitsa C, Field G D, Gauthier J L, Jepson L H, Sher A, Litke A M and Chichilnisky E J 2011 Correlated firing among major ganglion cell types in primate retina *J. Physiol.* **589** 75–86
- Grosberg L E *et al* 2017 Activation of ganglion cells and axon bundles using epiretinal electrical stimulation *J. Neurophysiol.* **118** 1457–71
- Habib A G, Cameron M A, Suaning G J, Lovell N H and Morley J W 2013 Spatially restricted electrical activation of retinal ganglion cells in the rabbit retina by hexapolar electrode return configuration *J. Neural. Eng.* **10** 036013
- Halupka K J, Shivdasani M N, Cloherty S L, Grayden D B, Wong Y T, Burkitt A N and Meffin H 2017 Prediction of cortical responses to simultaneous electrical stimulation of the retina *J. Neural. Eng.* **14** 016006
- Hashimoto T, Elder C M and Vitek J L 2002 A template subtraction method for stimulus artifact removal in high-frequency deep brain stimulation *J. Neurosci. Methods* **113** 181–6
- Heffer L F and Fallon J B 2008 A novel stimulus artifact removal technique for high-rate electrical stimulation *J. Neurosci. Methods* **170** 277–84
- Heitman A *et al* 2016 Testing pseudo-linear models of responses to natural scenes in primate retina (accepted) (<https://doi.org/10.1101/045336>)
- Ho E, Smith R, Goetz G, Lei X, Galambos L, Kamins T I, Harris J, Mathieson K, Palanker D and Sher A 2018 Spatiotemporal characteristics of retinal response to network-mediated photovoltaic stimulation *J. Neurophysiol.* **119** 389–400
- Hochstein S and Shapley R M 1976 Linear and nonlinear spatial subunits in Y cat retinal ganglion cells *J. Physiol.* **262** 265–84
- Hottoway P *et al* 2008 An integrated multichannel waveform generator for large-scale spatio-temporal stimulation of neural tissue *Analog Integr. Circuits Signal Process.* **55** 239–48
- Hottoway P, Skoczeń A, Gunning D E, Kachiguine S, Mathieson K, Sher A, Wiącek P, Litke A M and Dąbrowski W 2012 Properties and application of a multichannel integrated circuit for low-artifact, patterned electrical stimulation of neural tissue *J. Neural. Eng.* **9** 066005
- Humayun M S *et al* 2012 Interim results from the international trial of second sight's visual prosthesis *Ophthalmology* **119** 779–88
- Jarosiewicz B *et al* 2015 Virtual typing by people with tetraplegia using a self-calibrating intracortical brain-computer interface *Sci. Transl. Med.* **7** 313ra179
- Jepson L H *et al* 2013 Focal electrical stimulation of major ganglion cell types in the primate retina for the design of visual prostheses *J. Neurosci.* **33** 7194–205
- Jepson L H, Hottoway P, Mathieson K, Gunning D E, Dąbrowski W, Litke A M and Chichilnisky E J 2014a Spatially patterned electrical stimulation to enhance resolution of retinal prostheses *J. Neurosci.* **34** 4871–81
- Jepson L H, Hottoway P, Weiner G, Dabrowski W, Litke A and Chichilnisky E J 2014b High-fidelity reproduction of spatiotemporal visual signals for retinal prosthesis *Neuron* **83** 87–92
- Joarder S A, Abramian M, Suaning G J, Lovell N H and Dokos S 2011 A continuum model of retinal electrical stimulation *J. Neural. Eng.* **8** 066006

- Jones B W, Pfeiffer R L, Ferrell W D, Watt C B, Marmor M and Marc R E 2016 Retinal remodeling in human retinitis pigmentosa *Exp. Eye Res.* **150** 149–65
- Jun J J *et al* 2017 Fully integrated silicon probes for high-density recording of neural activity *Nature* **551** 232–6
- Kaardal J, Fitzgerald J D, Berry M J and Sharpee T O 2013 Identifying functional bases for multidimensional neural computations *Neural Comput.* **25** 1870–90
- Kandel E 2013 *Principles of Neural Science* 5th edn (New York: McGraw Hill)
- Kastner D B and Baccus S A 2013 Spatial segregation of adaptation and predictive sensitization in retinal ganglion cells *Neuron* **79** 541–54
- Kim K J and Rieke F 2001 Temporal contrast adaptation in the input and output signals of salamander retinal ganglion cells *J. Neurosci.* **21** 287–99
- Kling A *et al* 2020 Functional organization of midget and parasol ganglion cells in the human retina (<https://doi.org/10.1101/2020.08.07.240762>)
- Kowler E 2011 Eye movements: the past 25 years *Vision Res.* **51** 1457–83
- Krook-Magnuson E, Gelinas J N, Soltesz I and Buzsáki G 2015 Neuroelectronics and biooptics: closed-loop technologies in neurological disorders *JAMA Neurol.* **72** 823
- Lee B B, Martin P R and Grünert U 2010 Retinal connectivity and primate vision *Prog. Retin. Eye Res.* **29** 622–39
- Lee J-I and Im M 2019 Optimal electric stimulus amplitude improves the selectivity between responses of ON versus OFF types of retinal ganglion cells *IEEE Trans. Neural. Syst. Rehabil. Eng.* **27** 2015–24
- Lee S W, Eddington D K and Fried S I 2013 Responses to pulsatile subretinal electric stimulation: effects of amplitude and duration *J. Neurophysiol.* **109** 1954–68
- Li P H, Gauthier J L, Schiff M, Sher A, Ahn D, Field G D, Greschner M, Callaway E M, Litke A M and Chichilnisky E J 2015 Anatomical identification of extracellularly recorded cells in large-scale multielectrode recordings *J. Neurosci.* **35** 4663–75
- Litke A M *et al* 2004 What does the eye tell the brain?: development of a system for the large-scale recording of retinal output activity *IEEE Trans. Nucl. Sci.* **51** 1434–40
- Liu J K, Schreyer H M, Onken A, Rozenblit F, Khani M H, Krishnamoorthy V, Panzeri S and Gollisch T 2017 Inference of neuronal functional circuitry with spike-triggered non-negative matrix factorization *Nat. Commun.* **8** 149
- Lozano A M and Lipsman N 2013 Probing and regulating dysfunctional circuits using deep brain stimulation *Neuron* **77** 406–24
- Madugula S *et al* 2019 Using electrical images to predict electrical stimulation thresholds for epiretinal stimulation in *The Eye and the Chip (TEATC)*
- Madugula S *et al* 2020 Focal electrical stimulation of human retinal ganglion cells (submitted) (<https://doi.org/10.1101/2020.08.23.263608>)
- Maheswaranathan N *et al* 2019 The dynamic neural code of the retina for natural scenes (submitted) (<https://doi.org/10.1101/340943>)
- Maheswaranathan N, Kastner D B, Baccus S A and Ganguli S 2018 Inferring hidden structure in multilayered neural circuits *PLoS Comput. Biol.* **14** e1006291
- Marcus G and Koch C 2014 The future of brain implants *WSJ Online*, 14 March (available at: <https://www.wsj.com/articles/the-future-of-brain-implants-1394839583>) (Accessed: 2 April 2020)
- Martens H C F, Toader E, Decré M M J, Anderson D J, Vetter R, Kipke D R, Baker K B, Johnson M D and Vitek J L 2011 Spatial steering of deep brain stimulation volumes using a novel lead design *Clin. Neurophysiol.* **122** 558–66
- Mastroratte D N 1983 Correlated firing of cat retinal ganglion cells. I. Spontaneously active inputs to X- and Y-cells *J. Neurophysiol.* **49** 303–24
- Mathieson K *et al* 2012 Photovoltaic retinal prosthesis with high pixel density *Nat. Photonics* **6** 391–7
- Matteucci P B *et al* 2013 Current steering in retinal stimulation via a quasimonopolar stimulation paradigm *Invest. Ophthalmol. Visual Sci.* **54** 4307–20
- McFarland J M, Cui Y and Butts D A 2013 Inferring nonlinear neuronal computation based on physiologically plausible inputs *PLoS Comput. Biol.* **9** e1003143
- McIntosh L *et al* 2016 Deep learning models of the retinal response to natural scenes *Adv. Neural Inf. Process. Syst.* **29** 1369–77
- Mena G E, Grosberg L E, Madugula S, Hottoway P, Litke A, Cunningham J, Chichilnisky E J and Paninski L 2017 Electrical stimulus artifact cancellation and neural spike detection on large multi-electrode arrays *PLoS Comput. Biol.* **13** e1005842
- Meng K, Fellner A, Rattay F, Ghezzi D, Meffin H, Ibbotson M R and Kamenova T 2018 Upper stimulation threshold for retinal ganglion cell activation *J. Neural. Eng.* **15** 046012
- Muratore D G and Chichilnisky E J 2020 Artificial retina: a future cellular-resolution brain-machine interface *Chips 2030* (Berlin: Springer)
- Muratore D G, Tandon P, Wootters M, Chichilnisky E J, Mitra S and Murmann B 2019 A data-compressive wired-OR readout for massively parallel neural recording *IEEE Trans. Biomed. Circuits Syst.* **13** 1128–40
- Nanduri D 2011 Prosthetic vision in blind human patients: Predicting the percepts of epiretinal stimulation *PhD Thesis* (Los Angeles: University of Southern California)
- Nirenberg S and Pandarinath C 2012 Retinal prosthetic strategy with the capacity to restore normal vision *Proc. Natl Acad. Sci. USA* **109** 15012–7
- Obaid A *et al* 2020 Massively parallel microwire arrays integrated with CMOS chips for neural recording *Sci. Adv.* **6** eaay2789
- Palanker D 2014 Retinal prostheses for restoration of sight *Encycl. Comput. Neurosci.* 1–4
- Palanker D *et al* 2020 Photovoltaic restoration of central vision in atrophic age-related macular degeneration *Ophthalmology* **127** 1097–104
- Palanker D, Vankov A, Huie P and Baccus S 2005 Design of a high-resolution optoelectronic retinal prosthesis *J. Neural. Eng.* **2** S105–20
- Pandarinath C *et al* 2017 High performance communication by people with paralysis using an intracortical brain-computer interface *eLife* **6** e18554
- Parthasarathy N *et al* 2017 Neural networks for efficient bayesian decoding of natural images from retinal neurons in *Thirty-first Conf. on Neural Information Processing Systems* (<https://doi.org/10.1101/153759>)
- Pillow J W, Shlens J, Paninski L, Sher A, Litke A M, Chichilnisky E J and Simoncelli E P 2008 Spatio-temporal correlations and visual signalling in a complete neuronal population *Nature* **454** 995–9
- Rathbun D L, Ghorbani N, Shabani H, Zrenner E and Hosseinzadeh Z 2018 Spike-triggered average electrical stimuli as input filters for bionic vision—a perspective *J. Neural. Eng.* **15** 063002
- Ratnam K, Domdei N, Harmening W M and Roorda A 2017 Benefits of retinal image motion at the limits of spatial vision *J. Vis.* **17** 30
- Rattay F 1986 Analysis of models for external stimulation of axons *IEEE Trans. Biomed. Eng.* **33** 974–7
- Rattay F 1989 Analysis of models for extracellular fiber stimulation *IEEE Trans. Biomed. Eng.* **36** 676–82
- Rattay F 1999 The basic mechanism for the electrical stimulation of the nervous system *Neuroscience* **89** 335–46
- Rattay F, Paredes L P and Leao R N 2012 Strength–duration relationship for intra- versus extracellular stimulation with microelectrodes *Neuroscience* **214** 1–13
- Reinhard K and Münch T A 2019 What your eye tells your brain (submitted) (<https://doi.org/10.1101/766170>)

- Ren Y-M *et al* 2018 Changes in intrinsic excitability of ganglion cells in degenerated retinas of RCS rats *Int. J. Ophthalmol.* **11** 756–65
- Rhoades C E, Shah N P, Manookin M B, Brackbill N, Kling A, Goetz G, Sher A, Litke A M and Chichilnisky E J 2019 Unusual physiological properties of smooth monostriated ganglion cell types in primate retina *Neuron* **103** 658–672.e6
- Richard E, Goetz G A and Chichilnisky E J 2015 Recognizing retinal ganglion cells in the dark *Advances in Neural Information Processing Systems* pp 2476–84 (Montreal: Curran Associates, Inc.)
- Rieke F 1996 *Spikes: Exploring the Neural Code* (Cambridge, MA: MIT Press)
- Rodieck R W 1998 *The First Steps in Seeing* (Oxford: Oxford University Press)
- Sabbah N, Authie C N, Sanda N, Mohand-Said S, Sahel J-A and Safran A B 2014 Importance of eye position on spatial localization in blind subjects wearing an Argus II retinal prosthesis *Invest. Ophthalmol. Vis. Sci.* **55** 8259–66
- Salisbury J M and Palmer S E 2016 Optimal prediction in the retina and natural motion statistics *J. Stat. Phys.* **162** 1309–23
- Segal I Y, Giladi C, Gedalin M, Rucci M, Ben-Tov M, Kushinsky Y, Mokeichev A and Segev R 2015 Decorrelation of retinal response to natural scenes by fixational eye movements *Proc. Natl Acad. Sci. USA* **112** 3110–5
- Sekirnjak C *et al* 2007 Electrical stimulation of mammalian retinal ganglion cells using dense arrays of small-diameter electrodes *Artif. Sight* pp 333–45
- Sekirnjak C, Hottowy P, Sher A, Dabrowski W, Litke A M and Chichilnisky E J 2008 High-resolution electrical stimulation of primate retina for epiretinal implant design *J. Neurosci.* **28** 4446–56
- Sekirnjak C, Jepson L H, Hottowy P, Sher A, Dabrowski W, Litke A M and Chichilnisky E J 2011 Changes in physiological properties of rat ganglion cells during retinal degeneration *J. Neurophysiol.* **105** 2560–71
- Shah N P *et al* 2018 Learning a neural response metric for retinal prosthesis in *Int. Conf. on Learning Representations* (<https://doi.org/10.1101/226530>)
- Shah N P *et al* 2019a Optimization of electrical stimulation for a high-fidelity artificial retina. 2019 9th Int. IEEE/EMBS Conf. on Neural Engineering (NER) pp 714–8
- Shah N P *et al* 2019b Learning variability in the neural code of the retina *Computational and Systems Neuroscience (Cosyne)* (Lisbon: Cosyne)
- Shah N P *et al* 2020 Inference of nonlinear receptive field subunits with spike-triggered clustering *eLife* **9** e45743
- Shannon R V 2012 Advances in auditory prostheses *Curr. Opin. Neurol.* **25** 61–66
- Sinclair N C, Shivdasani M N, Perera T, Gillespie L N, Mcdermott H J, Ayton L N and Blamey P J 2016 The appearance of phosphenes elicited using a suprachoroidal retinal prosthesis *Invest. Ophthalmol. Vis. Sci.* **57** 4948–61
- Sinha R, Hoon M, Baudin J, Okawa H, Wong R O L and Rieke F 2017 Cellular and circuit mechanisms shaping the perceptual properties of the primate fovea *Cell* **168** 413–426.e12
- Smirnakis S M, Berry M J, Warland D K, Bialek W and Meister M 1997 Adaptation of retinal processing to image contrast and spatial scale *Nature* **386** 69–73
- Soto F *et al* 2020 Efficient coding by midget and parasol ganglion cells in the human retina *Neuron* **107** 656–66
- Spencer T C *et al* 2018 Electrical field shaping techniques in a feline model of retinal degeneration. *Annual Int. Conf. of the IEEE Eng. Med. Biol. Soc.* pp 1222–5
- Spencer T C, Fallon J B and Shivdasani M N 2018 Creating virtual electrodes with 2D current steering *J. Neural. Eng.* **15** 035002
- Spencer T C, Fallon J B, Thien P C and Shivdasani M N 2016 Spatial restriction of neural activation using focused multipolar stimulation with a retinal prosthesis *Invest. Ophthalmol. Vis. Sci.* **57** 3181–91
- Stanley G B, Li F F and Dan Y 1999 Reconstruction of natural scenes from ensemble responses in the lateral geniculate nucleus *J. Neurosci.* **19** 8036–42
- Stavisky S D *et al* 2019 Neural ensemble dynamics in dorsal motor cortex during speech in people with paralysis *eLife* **8** e46015
- Stingl K *et al* 2013 Artificial vision with wirelessly powered subretinal electronic implant alpha-IMS *Proc. Biol. Sci.* **280** 20130077
- Su C-Y, Menuz K and Carlson J R 2009 Olfactory perception: receptors, cells, and circuits *Cell* **139** 45–59
- Sweeney J D, Ksienski D A and Mortimer J T 1990 A nerve cuff technique for selective excitation of peripheral nerve trunk regions *IEEE Trans. Biomed. Eng.* **37** 706–15
- Tandon P *et al* 2017 Automatic identification and avoidance of axon bundle activation for epiretinal prosthesis in *The Eye and the Chip (TEATC)*
- Townshend B and White R L 1987 Reduction of electrical interaction in auditory prostheses *IEEE Trans. Biomed. Eng.* **34** 891–7
- Trenholm S and Awatramani G B 2015 Origins of spontaneous activity in the degenerating retina *Frontiers Cell Neurosci.* **9** 277
- Turner M H and Rieke F 2016 Synaptic rectification controls nonlinear spatial integration of natural visual inputs *Neuron* **90** 1257–71
- Turner M H, Sanchez Giraldo L G, Schwartz O and Rieke F 2019 Stimulus- and goal-oriented frameworks for understanding natural vision *Nat. Neurosci.* **22** 15–24
- Tyler D J 2015 Neural interfaces for somatosensory feedback: bringing life to a prosthesis *Curr. Opin. Neurol.* **28** 574–81
- van Rossum M C W 2001 A novel spike distance *Neural Comput.* **13** 751–63
- Victor J D 2005 Spike train metrics *Curr. Opin. Neurobiol.* **15** 585–92
- Victor J D and Purpura K P 1996 Nature and precision of temporal coding in visual cortex: a metric-space analysis *J. Neurophysiol.* **76** 1310–26
- Victor J D and Purpura K P 1997 Metric-space analysis of spike trains: theory, algorithms and application *Network: Comput. Neural Syst.* **8** 127–64
- Wachtler T, Wehrhahn C and Lee B B 1996 A simple model of human foveal ganglion cell responses to hyperacuity stimuli *J. Comput. Neurosci.* **3** 73–82
- Wagenaar D A and Potter S M 2002 Real-time multi-channel stimulus artifact suppression by local curve fitting *J. Neurosci. Methods* **120** 113–20
- Wandell B A 1995 *Foundations of Vision* (Oxford: Sinauer Associates Incorporated)
- Wang Z, Bovik A C, Sheikh H R and Simoncelli E P 2004 Image quality assessment: from error visibility to structural similarity *IEEE Trans. Image Process.* **13** 600–12
- Warland D K, Reinagel P and Meister M 1997 Decoding visual information from a population of retinal ganglion cells *J. Neurophysiol.* **78** 2336–50
- Weber A 1997 The USC-SIPI image database Signal and Image Processing Institute of the University of Southern California California (<http://sipi.usc.edu/database/database.php>)
- Weitz A C, Nanduri D, Behrend M R, Gonzalez-Calle A, Greenberg R J, Humayun M S, Chow R H and Weiland J D 2015 Improving the spatial resolution of epiretinal implants by increasing stimulus pulse duration *Sci. Transl. Med.* **7** 318ra203
- Wichmann T and Devergnas A 2011 A novel device to suppress electrical stimulus artifacts in electrophysiological experiments *J. Neurosci. Methods* **201** 1–8
- Wong Y T *et al* 2009 Focal activation of the feline retina via a suprachoroidal electrode array *Vision Res.* **49** 825–33
- Zhang Y, Jia S, Zheng Y, Yu Z, Tian Y, Ma S, Huang T and Liu J K 2020 Reconstruction of natural visual scenes from neural spikes with deep neural networks *Neural Netw.* **125** 19–30