

Review



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Genetic dissection of neural circuits underlying sexually dimorphic social behaviours

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The unique hormonal, genetic and epigenetic environments of males and females during development and adulthood shape the neural circuitry of the brain. These differences in neural circuitry result in sex-typical displays of social behaviours such as mating and aggression. Like other neural circuits, those underlying sex-typical social behaviours weave through complex brain regions that control a variety of diverse behaviours. For this reason, the functional dissection of neural circuits underlying sex-typical social behaviours has proved to be difficult. However, molecularly discrete neuronal subpopulations can be identified in the heterogeneous brain regions that control sex-typical social behaviours. In addition, the actions of oestrogens and androgens produce sex differences in gene expression within these brain regions, thereby highlighting the neuronal subpopulations most likely to control sexually dimorphic social behaviours. These conditions permit the implementation of innovative genetic approaches that, in mammals, are most highly advanced in the laboratory mouse. Such approaches have greatly advanced our understanding of the functional significance of sexually dimorphic neural circuits in the brain. In this review, we discuss the neural circuitry of sex-typical social behaviours in mice while highlighting the genetic technical innovations that have advanced the field.

1. Introduction

Sexually reproducing animals exhibit sex-typical displays of social behaviours, such as mating and aggression. Such sexual dimorphisms in behaviour can be qualitative or quantitative in nature, and they arise from sexually differentiated neural circuits, which in turn are shaped by the varying hormonal, genetic and epigenetic environments of males and females during development and adulthood [1–6]. Hormones such as oestrogens and androgens exert their effects by binding to their respective membrane-bound and nuclear receptors [7–8]. Nuclear oestrogen and androgen receptors (ARs) are transcription factors that can bind to hormone-responsive DNA elements to regulate expression of target genes [6,9], and, in neurons, presumably lead to changes in neuronal connectivity and excitability and ultimately, behaviour.

Many sex differences in gene expression have been identified [10–18], and hormone-driven sex differences in gene expression highlight neuronal subpopulations within the brain areas implicated in sex-typical social behaviours that are most likely to control these behaviours. This information is useful because the neural circuits underlying these behaviours, like many neural circuits, are embedded in brain regions comprising a tangled mixture of neurons that control diverse behaviours. For example, many studies implicate the bed nucleus of the stria terminalis (BNST) in social behaviours [19–26]. However, the BNST is also implicated in other behaviours such as stress responses, reward responses and salt intake [27–35]. With traditional approaches, it is not possible to prospectively identify or manipulate the relevant subpopulations within the BNST that control sex-typical social behaviours. Hormone-dependent gene expression patterns define specific pools of neurons in the BNST as well as other heterogeneous brain regions that control

sex-typical social behaviours [10,36–40]. The use of genetic strategies to selectively study and manipulate such molecularly defined neuronal subpopulations within the BNST and elsewhere will be essential in the attempt to untangle and understand the neural circuits underlying sexually dimorphic behaviour.

This review provides an overview of our current understanding of the neural circuits underlying sexually dimorphic social behaviours while highlighting genetic approaches that continue to advance the field. Important contributions to our understanding of the neural circuitry of sex-typical social behaviours have also been made by work on other organisms, including humans (e.g. [41–53]). However, in mammals, state-of-the-art genetic tools are currently most powerfully implemented using the laboratory mouse, and therefore, this review will focus on the neural pathways underlying social interactions in rodents. We discuss the role of sensory input in the control of social behaviours, the brain regions implicated in social behaviours, the behavioural significance of molecularly defined neuronal pools within these brain areas and future directions for the study of sexually dimorphic social behaviours.

2. Sensory input driving sexually dimorphic social behaviours

Sex-typical social behaviours occur in response to sensory cues from the environment [54–62]. The predominant sensory cues that trigger these behaviours, in mice and many other animals, are pheromones detected by chemosensory neurons in two epithelia in the nose: the main olfactory epithelium (MOE) and the vomeronasal organ (VNO) [63]. Neurons in the MOE and VNO express G-protein-coupled chemosensory receptors from unrelated gene families [64] and detect volatile and contact-based pheromones, respectively [65–66]. The importance of pheromone signalling via the VNO for the control of sex-typical social behaviours has long been established by traditional lesioning studies [67]. However, implementation of genetic approaches has revealed previously unappreciated complexity of the chemosensory control of these behaviours [54–62].

The generation of genetically modified mice bearing a loss of function for specific genes has been a fundamental tool in the dissection of the neural circuits underlying sex-typical social behaviours. These techniques will continue to be a valuable resource for research as recent advances in the application of CRISPR-Cas systems for gene editing have greatly increased the speed and efficiency of generating genetic knockouts [68–70]. The use of targeted gene knockouts has made it possible to study the contributions of contact-based VNO or volatile MOE sensory input for the display of sex-typical social behaviours by genetically disabling one signalling pathway while keeping the other functionally intact. These studies conducted in many laboratories, including ours, reveal that mating and aggression behaviours require the coordinated function of both epithelia [54–61].

Targeted deletion of Trpc2, a cation channel expressed in the VNO but not MOE, results in mice with a functionally disabled VNO ([58,60], but see [71,72]). Male Trpc2 knockout mice exhibit male-typical mounting and copulatory behaviours towards not only females but also other males, and male-typical aggressive attacks are abrogated in these mice

[58,60]. These findings suggest that contact-based male pheromones detected by the VNO normally inhibit mating and trigger aggression towards other males. Interestingly, female Trpc2 knockout mice also display high levels of male-typical mounting behaviour towards mice of either sex [57], suggesting that the neural circuit for male-typical mating behaviour is present in both sexes but is normally inhibited by sensory input from the VNO. The presence of a shared male-type mating circuitry in both sexes is consistent with prior classical endocrinological studies showing that adult females treated with testosterone will display male-typical mating behaviour [73]. Therefore, circulating testosterone can override the contact pheromone-based VNO-mediated sensory inhibition of male-typical mating behaviours. The importance of the detection of volatile pheromones by the MOE has been uncovered using mice with a targeted deletion of Cnga2, a cation channel expressed in the MOE but not VNO, leading to disabled MOE signalling. Male Cnga2 knockout mice exhibit a profound reduction in male-typical mounting and copulatory behaviours as well as male-typical aggressive attacks [59,61,74]. Thus, both male-typical mating and aggression behaviours require sensory input from the MOE. Studies using these mutants also demonstrate that the VNO and MOE are required for female-typical displays of sexually dimorphic social behaviours, such as sexual receptivity and maternal aggression [54].

Taken together, these genetic studies suggest a model in which sex differences in neural circuits underlying sexually dimorphic social interactions regulate the display of these behaviours such that contact-based VNO sensory input decreases and testosterone increases the probability of displaying male-typical mating, whereas both contact-based VNO and volatile MOE sensory input increase the probability of displaying male-typical aggression. Therefore, it appears that male-typical mating behaviours are normally inhibited in adult females by a functional VNO and the absence of high, male-typical levels of testosterone. Female mice rarely attack males unless nursing, and there appears to be no qualitative sex difference in the expression of the chemoreceptors [65]. Therefore, the lack of male-typical aggression in females is most likely regulated by a sexual dimorphism downstream of chemosensory input in the underlying neural circuit.

3. Neural circuits underlying sexually dimorphic social behaviours

(a) Overview of brain regions implicated in sexually dimorphic social behaviours

The sensory inputs of the VNO and MOE maintain their anatomic segregation with central projections to the accessory olfactory bulb (AOB) and the main olfactory bulb (MOB), respectively [63,66]. AOB projection neurons send axons to the medial amygdala (MeA) as well as the posteromedial cortical amygdala [75–77]. By contrast, MOB projection neurons send axons to several cortical regions, including the postero-lateral cortical amygdala [76,78–80]. As discussed above, both the VNO and MOE regulate sex-typical social behaviours, indicating an interaction between or convergence of these two chemosensory circuits. Circuit mapping reveals a convergence of the connections of the projection targets of the AOB and MOB in the BNST and several hypothalamic

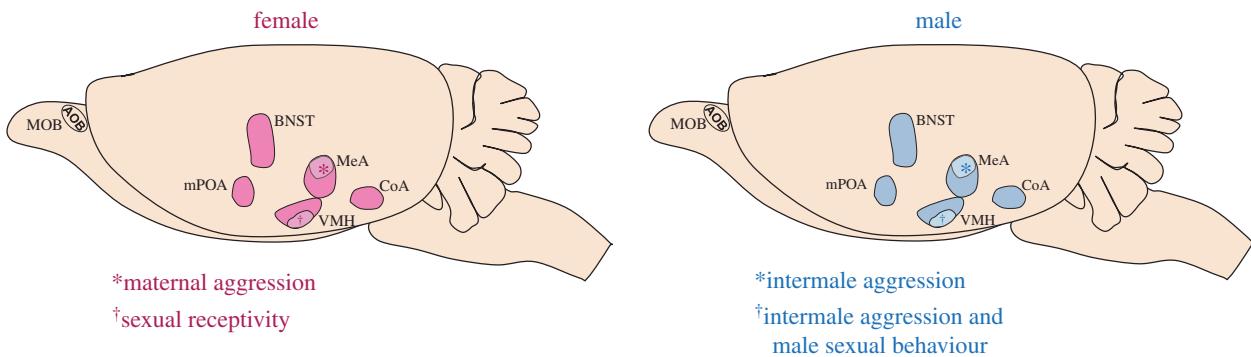


Figure 1. Brain regions implicated in sexually dimorphic social behaviours. A shared set of brain regions has been implicated in the control of sexually dimorphic social behaviours in females and males, raising the issue of how neurons in these areas control distinct behaviours in the two sexes. A related issue is that the molecular identity of neurons in these functionally and molecularly heterogeneous brain regions is largely unknown. Recent molecular genetic studies have now identified discrete subsets of neurons, representing 10–20% of neurons in these regions, that control one or a few sexually dimorphic social behaviours but not other behaviours regulated by that entire brain region. Thus, the control of complex social behaviours appears to be modularly organized at the level of molecularly defined neurons. Another general principle emerging from these studies is that these neurons are functionally bivalent in the sense that they control distinct behaviours in both sexes. Such sexually dimorphic function is likely to be regulated by sex differences in gene expression or connectivity. Indeed, both the MeA and VMH express genes in a sexually dimorphic manner (e.g. [10,36,37]), and a recent study showed that VMH neurons regulating social behaviours project in a sex-typical manner [37]. AOB, accessory olfactory bulb; CoA, cortical amygdala encompassing both the posterolateral and posteromedial components that receive inputs from the MOB and AOB, respectively; MOB, main olfactory bulb. *, aromatase-expressing neurons in the posterodorsal MeA (MeApd) control maternal aggression and intermale aggression; †, progesterone receptor (PR)-expressing neurons in the ventrolateral subdivision of VMH (VMHvl) control mating behaviour in both sexes and aggression in males. (Online version in colour.)

nuclei [81–85]. In addition, some, but not all, studies report MOB projections directly to the MeA, thereby providing another site of convergence of the VNO and MOE pathways [78,80,83,86]. Neurons in the MeA, in turn, project to targets including the ventromedial hypothalamus (VMH), the BNST and the medial preoptic area (mPOA). These regions are interconnected, and decades of lesion or stimulation studies of these regions have revealed their importance in the control of mating and aggression behaviours in both sexes [26–27,87–98]. How molecularly defined neurons within these brain regions interact to produce sex-typical behaviours, and whether these interconnected pathways comprise a single neural circuit or a set of separate neural circuits that control sex-typical mating and aggression behaviours, is currently unclear and under investigation (figure 1).

(b) Sex differences in gene expression

The MeA, VMH, BNST and mPOA exhibit sex differences such that the number of neurons or innervation within these regions are different in males and females [10,36–37,40,99–101], and with few exceptions, subsets of adult neurons within these regions express one or more sex hormone receptors [10,36–37,40,102–104]. As discussed above, these receptors can regulate gene expression, and many hormone-dependent sex differences in gene expression have been identified, including in the sex hormone receptors themselves and the enzyme aromatase that converts testosterone into 17 β -oestradiol [10–18,36–37,40]. For example, we have used genetic strategies to visualize the expression pattern of aromatase in the brain and discovered that it is expressed in a few discrete areas, including the posterodorsal MeA (MeApd) and the posteromedial part of medial division of the BNST (BNSTmpm). Aromatase-expressing neurons in the MeApd and BNSTmpm exhibit sex differences in cell number, and this sexual dimorphism is masculinized by neonatal oestrogen exposure [36]. Notably, this masculinization of the number of aromatase-expressing neurons is accompanied by a masculinization

of aggression behaviour in females even in the absence of adult supplementation of testosterone.

Numerous, novel sex differences in gene expression patterns within the MeA, mPOA, VMH and BNST have been identified [10]. Using genome-wide expression profiling in conjunction with *in situ* hybridization, we discovered that the regulation of sexually dimorphic gene expression patterns is complex as many individual genes are upregulated in different brain regions in the two sexes [10]. Consistent with this complexity, we showed that in males many, but not all, of the sex differences in gene expression are dependent upon adult testosterone. However, in females, the sex differences in gene expression are mostly independent of adult ovarian hormones, suggesting a developmental pre-patterning of the dimorphic transcriptional programme in females. Moreover, we demonstrated that these sexually dimorphic genes play important, modular roles in sexually dimorphic social behaviours. Mice singly homozygous null for these genes exhibit specific deficits in one or more components of male or female mating behaviour (Brs3, Syt4 and Cckar), male-typical aggression (Brs3) or maternal care (Irs4), while other sex-typical behaviours remain unaffected. Each of these dimorphic genes is expressed in specific neurons within the MeA, VMH or other areas, implicating one or more neuronal pools in distinct components of sex-typical social behaviours. These findings indicate that sex-typical social behaviours are modular in that specific components of behaviour (such as male-typical aggressive attacks and male-typical marking behaviour) are controlled by distinct sets of genes. For example, Cckar, a G-protein-coupled receptor for the neuropeptide cholecystokinin, is expressed in a subset of neurons within the VMH, and Cckar is required for female sexual receptivity but not for maternal behaviours and the oestrous cycle [10]. Thus, unlike castrates or animals mutant for sex hormone receptors that exhibit global deficits in sex-typical behaviours, mice mutant for genes that are downstream of sex hormone signalling exhibit phenotypes restricted to specific components

of sexually dimorphic behaviours. These findings suggest a model in which the components of individual dimorphic behaviours are controlled in a modular manner by genetically separable pathways comprising sexually dimorphic molecularly defined neuronal subpopulations.

(c) Genetic dissection of molecularly defined neuronal populations underlying sexually dimorphic behaviours

The utilization of conditional genetic approaches such as the Cre-lox system to study molecularly defined sexually dimorphic neuronal subpopulations has greatly advanced our understanding of the neural circuits underlying sex-typical social behaviours. The MeA, VMH, BNST and mPOA each contains a complex mixture of molecularly discrete populations of neurons intermingled with each other [10,36–37,40, 105–106]. Therefore, the neural pathways underlying sex-typical social behaviours are embedded within an assortment of neural circuits involved in many unrelated behaviours. The use of conditional genetic approaches is necessary to study the neurons relevant for social behaviours in isolation from their functionally distinct neighbours [107–109]. The use of mice genetically modified to express Cre in molecularly defined neuronal subpopulations of neurons in conjunction with virally encoded Cre-dependent transgenes has permitted projection-mapping as well as functional characterization of these neurons. The viruses used in such studies are stereotactically delivered to the region of interest where they indiscriminately infect all neurons, but the virally encoded transgene is expressed only in neurons that express Cre.

We recently used this genetic approach to study the role of the MeA in sex-typical social behaviours [38]. The MeA is a large complex that controls many diverse behaviours, including dimorphic patterns of mating and aggression, social memory, stress, anxiety and defensive response to predators [38,110–113]. The functional diversity of the MeA is reflected in its molecular heterogeneity and the diversity of its projections [10,105,114–116]. It is not clear how the MeA integrates sensory and physiological cues to regulate such an array of diverse behaviours. We have discovered a sexually dimorphic subpopulation of neurons in the MeApd that expresses the enzyme aromatase such that there are more aromatase-expressing neurons in males compared with females [36]. Aromatase itself is required for the normal display of male-typical mating and aggression behaviours [117–120]. To study the functional significance of the aromatase-expressing neuronal subpopulation within the MeApd, we specifically ablated these neurons in adult mice [38], using a virally encoded Cre-dependent designer executioner caspase-3 [37,121]. These studies were done in genetically modified mice that we generated in order to express Cre in aromatase-expressing cells (*aro*^{cre} mice). Targeted delivery of the virally encoded caspase-3 to the MeA specifically induced apoptosis of aromatase-expressing MeApd neurons in *aro*^{cre} but not control wild-type mice. Importantly, this caspase-3-induced apoptosis is restricted to Cre-expressing neurons and spares neighbouring neurons that do not express aromatase (Cre) [37–38,121]. Ablation of these neurons in males significantly reduced specific components of male-typical aggression while male-typical mating behaviours were unaffected, whereas ablation of these

neurons in females significantly reduced specific components of maternal aggression while maternal care and female-typical mating behaviours were unaffected. Furthermore, using this approach to exclusively express DREADD/Gi, an engineered inhibitory G-protein-coupled receptor activated by a heterologous ligand [122], in aromatase-expressing MeApd neurons, we found that pharmacogenetic inhibition of aromatase-expressing MeApd neurons recapitulated these behavioural deficits in both sexes. The sensory, hormonal and contextual requirements for male-typical and maternal aggression are very different [123–125]. Therefore, aromatase-expressing MeApd neurons appear to be functionally bivalent in that they control distinct forms of aggression in each sex. In addition, our findings demonstrate that it is possible to dissociate specific components of a complex social display such as aggression without altering other features of social interactions. Indeed, our manipulations show that it is possible to dissociate maternal care of pups from maternal defence of pups from intruders. The entire MeA is critical for aggression as well as mating behaviours in both sexes [105–106,126–129], and a recent study described the important contributions of GABAergic MeApd neurons to both male-typical mating and aggression behaviours [113]. Importantly, aromatase-expressing neurons are a subset of the population of GABAergic neurons within the MeApd [38]. Therefore, by studying a small molecularly defined subpopulation of neurons expressing a gene (aromatase) functionally important for social behaviours, we have identified a subset of neurons within the MeApd that specifically regulate intermale and maternal aggression.

For almost a century, experiments have implicated the VMH or adjacent hypothalamic regions in modulating aggression behaviour in diverse species [93,130–131]. Only recently has the ventrolateral VMH (VMHvl) been implicated as a key aggression-eliciting centre in the male mouse [94]. *In vivo* recordings in freely moving male mice reveal an increase in VMHvl activity during male-typical aggression, and optogenetic stimulation of VMHvl neurons elicits indiscriminate aggressive attacks towards both males and females. The VMH is molecularly heterogeneous and regulates both female sexual receptivity and male-typical aggression behaviours as well as defensive reactions to predators and energy balance in diverse animals, including humans [89,93–97,130,132–146]. Lesions and other manipulations that do not target molecularly defined neurons within the VMH can yield conflicting behavioural phenotypes [96–97,135,137], perhaps owing to non-targeted manipulation of these heterogeneous neurons or destruction of fibres of passage within this region. Indeed, response profiles of the neural activity of VMHvl neurons are complex and heterogeneous, with some subpopulations responding maximally during male-typical aggression behaviour and others responding maximally during other behaviours [147]. To identify the VMH neurons that specifically regulate sex-typical social behaviours, we used the genetic strategy discussed above to ablate progesterone receptor (PR)-expressing neurons in the VMHvl of adult mice that we genetically modified to express Cre in PR-expressing cells [37]. PR-expressing VMHvl neurons are a subset of neurons within the VMHvl, which itself represents a subset of neurons within the VMH. PR expression is sexually dimorphic such that there are more PR-expressing VMHvl neurons in females than males. We discovered that ablation of these PR-expressing neurons in the adult female VMHvl causes a profound reduction in sexual receptivity, and ablation of the

corresponding neurons in males causes a significant reduction in both male-typical mating and aggression behaviour. Thus, these PR-expressing VMHvl neurons, like aromatase-expressing MeApd neurons, are functionally bivalent in that they regulate distinct dimorphic behaviours in the two sexes (i.e. sexual receptivity in females and male-typical mating and aggression in males).

These studies in the MeA and VMH therefore suggest a principle whereby a common pool of molecularly defined neurons is present in both sexes, but sex differences in gene expression or the neural circuit within which these neurons are embedded enable these pathways to transform sensory input into sexually dimorphic behaviours [6]. In both instances, characterization of neurons expressing a functionally important gene (aromatase, PR) reveals that these neurons regulate specific aspects of sexually dimorphic behaviour, whereas the genes themselves are expressed in multiple brain regions and regulate multiple aspects of these behaviours. Moreover, PR-expressing neurons in the VMHvl also express Cckar in a sexually dimorphic manner [10] such that there is little Cckar expression in males in this region, and its expression peaks in females during oestrus, a period of maximal sexual receptivity. As discussed earlier, females null for Cckar have a significant and specific reduction in sexual receptivity. Strikingly, ablation of PR- and Cckar-expressing VMHvl neurons leads to a correspondingly profound and specific diminution in sexual receptivity [37]. We therefore anticipate that other genes whose expression is regulated by sex hormone signalling (e.g. [10]) will similarly highlight neuronal subsets that also regulate sexually dimorphic behaviour in a modular manner.

How PR-expressing VMHvl neurons influence both male-typical mating and aggression behaviour is unclear. In one scenario, the same neurons mediate both male-typical mating and aggression behaviour via distinct patterns of neural activity. A recent study provides support for this scenario [148]. The vast majority of PR-expressing neurons in the VMHvl also express oestrogen receptor α (ER α or Esr1) [37]. Optogenetic stimulation of ER α -expressing VMHvl neurons elicits male-typical aggressive attacks in male mice [148]. Surprisingly, weaker optogenetic stimulation of these neurons elicits male-typical mating behaviour, rather than aggression, towards both females and males. These findings suggest that changes in neural activity in the VMHvl resulting in an increase in the number of active neurons or the average level of activity per neuron promotes a transition from male-typical mating to aggression behaviour. Alternatively, the subpopulation of ER α -expressing neurons in the VMHvl could consist of at least two molecularly distinct neuronal subsets, with one regulating male-typical mating and the other aggression. It is possible that different intensities of optogenetic stimulation preferentially recruit one or the other population to drive mating or aggression. Future studies manipulating the activity of the specific projections of molecularly defined neuronal subsets to identified target regions will greatly inform our understanding of the functional connectivity of the neural circuitry underlying sex-typical social behaviours.

4. Future directions of the study of sexually dimorphic social behaviours

A more mechanistic understanding of the neural circuits underlying sex-typical social behaviours will require a

comprehensive inter-disciplinary effort using innovative genetic strategies to map the anterograde and retrograde connectivity of molecularly defined subpopulations of neurons, followed by the optogenetic manipulation of specific projection targets and recording of the activity patterns *in vivo* of those same molecularly defined subpopulations during sex-typical social behaviours. Such anatomic circuit mapping endeavours will rely on recent advances in optogenetics as well as virally encoded trans-synaptic tracers [149–153] to facilitate the characterization of the neural connectivity of genetically defined pools of neurons. In addition to anatomic and optogenetic circuit mapping, it will be critical to use recording or imaging approaches to define activity patterns in molecularly defined neuronal populations. Recent advances in genetically encoded calcium sensors [154–155] in conjunction with advances in *in vivo* imaging via fibre optic cables or microscopy [156–159] will enable the recording of neural activity of genetically defined subsets of neurons during the display of sex-typical social behaviours. Such studies will provide insights into the information processed by molecularly defined neuronal pools at different levels in the neural circuits underlying sex-typical social behaviours.

In addition to an anatomic and functional delineation of neural circuits underlying sex-typical social behaviours, it will be important to understand the role of sexually dimorphic gene expression patterns in these behaviours. The use of conditional gene knockouts will address issues relating to the role of the gene in the development and function of the neural circuit, and it will also serve to localize gene action to specific neuronal populations that regulate the behaviour. For example, we and others have studied the functional role of the AR in the brain in male-typical social behaviours [39,160]. Constitutive, systemic deletion of AR leads to feminized external physical characteristics and demasculinized behaviours (*tfm* mice). However, these *tfm* males undergo testicular atrophy shortly after the critical period, leaving open the question of whether testosterone signalling via AR is required perinatally to masculinize the brain in an organizational manner. Alternatively, early masculinization of the brain may be solely regulated by oestrogens that are synthesized locally via aromatization of circulating testosterone. Targeted deletion of AR in the nervous system has shed light on this important question. Male mice in which AR is deleted in neural precursors but unaltered in other cells before the organizational critical period are externally masculinized and have physiological levels of circulating testosterone. Moreover, these mutant males exhibit qualitatively male-typical displays of social behaviours with a significant, quantitative reduction in various aspects of reproductive and territorial behaviours [39]. Taken together with classic endocrinological and other genetic studies, the results from a nervous system-restricted deletion of AR show that a functional AR is not required to masculinize the brain during the organizational period, but it is required for the normal intensity of male-typical social behaviours in adult life. In future studies, it will be important to use intersectional genetic strategies, with mice expressing Cre in specific neurons or targeted delivery of virally encoded Cre, to further localize the requirement of AR in specific neuronal subsets for the normal intensity of male-typical behavioural displays. The spatial and temporal disruption of other sexually dimorphically expressed genes will similarly

provide important insights into the molecular control of neural circuits underlying these social behaviours.

5. Conclusion

Outstanding progress has been made in our understanding of how the brain generates sexually dimorphic behaviours, and the application of the innovative genetic strategies discussed in this review will provide important new mechanistic insights into the molecular and neural control of sex-typical behaviours. The modular or specialized function of discrete molecularly defined neuronal populations uncovered by recent studies is a common theme emerging from the genetic studies of sex-typical social interactions in mice [6]. It will be important to test whether other complex behaviours are regulated in a similarly modular manner at the level of genes and neurons.

Healthy social interactions are an essential component of well-being and personal and professional success, and deficits in social interactions contribute to the emotional and

financial burden of many neurological and psychiatric disorders such as Alzheimer's disease and autism spectrum disorders. Furthermore, many neuro-psychiatric disorders exhibit sex differences in incidence, prevalence or disease outcome [161–165]. The reasons underlying these sex differences are poorly understood. We anticipate that insights from studies in mice using state-of-the-art genetic tools will reveal how sex differences in the brain translate into sexual dimorphisms in social behaviour in health and also render the brain differentially susceptible to disease between men and women.

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References

- Phoenix CH, Goy RW, Gerall AA, Young WC. 1959 Organizing action of prenatally administered testosterone propionate on the tissues mediating mating behavior in the female guinea pig. *Endocrinology* **65**, 369–382. (doi:10.1016/j.yhbeh.2009.03.015)
- McCarthy MM. 1994 Molecular aspects of sexual differentiation of the rodent brain. *Psychoneuroendocrinology* **19**, 415–427. (doi:10.1016/0306-4530(94)90029-9)
- Arnold AP. 2009 The organizational–activational hypothesis as the foundation for a unified theory of sexual differentiation of all mammalian tissues. *Horm. Behav.* **55**, 570–578. (doi:10.1016/j.yhbeh.2009.03.011)
- McCarthy MM. 2009 The epigenetics of sex differences in the brain. *J. Neurosci.* **29**, 12 815–12 823. (doi:10.1523/JNEUROSCI.3331-09.2009)
- McCarthy MM, Arnold AP. 2011 Reframing sexual differentiation of the brain. *Nat. Neurosci.* **14**, 677–683. (doi:10.1038/nn.2834)
- Yang CF, Shah NM. 2014 Representing sex in the brain, one module at a time. *Neuron* **82**, 261–278. (doi:10.1016/j.neuron.2014.03.029)
- Mangelsdorf DJ *et al.* 1995 The nuclear receptor superfamily: the second decade. *Cell* **83**, 835–839. (doi:10.1016/0092-8674(95)90199-X)
- Ervin KS, Lymer JM, Matta R, Clipperton-Allen AE, Kavaliers M, Choleris E. 2015 Estrogen involvement in social behavior in rodents: rapid and long-term actions. *Horm. Behav.* **74**, 53–76. (doi:10.1016/j.yhbeh.2015.05.023)
- Beato M, Sánchez-Pacheco A. 1996 Interaction of steroid hormone receptors with the transcription initiation complex. *Endocr. Rev.* **17**, 587–609. (doi:10.1210/edrv-17-6-587)
- Xu X, Coats JK, Yang CF, Wang A, Ahmed OM, Alvarado M, Izumi T, Shah NM. 2012 Modular genetic control of sexually dimorphic behaviors. *Cell* **148**, 596–607. (doi:10.1016/j.cell.2011.12.018)
- Dewing P, Shi T, Horvath S, Vilain E. 2003 Sexually dimorphic gene expression in mouse brain precedes gonadal differentiation. *Brain Res. Mol. Brain Res.* **118**, 82–90. (doi:10.1016/S0169-328X(03)00339-5)
- Dewing P *et al.* 2006 Direct regulation of adult brain function by the male-specific factor SRY. *Curr. Biol.* **16**, 415–420. (doi:10.1016/j.cub.2006.01.017)
- Edelmann M, Wolfe C, Scordalakes EM, Rissman EF, Tobet S. 2007 Neuronal nitric oxide synthase and calbindin delineate sex differences in the developing hypothalamus and preoptic area. *Dev. Neurobiol.* **67**, 1371–1381. (doi:10.1002/dneu.20507)
- Gagnidze K, Pfaff DW, Mong JA. 2010 Gene expression in neuroendocrine cells during the critical period for sexual differentiation of the brain. *Prog. Brain Res.* **186**, 97–111. (doi:10.1016/B978-0-444-53630-3.00007-5)
- Rinn JL, Rozowsky JS, Laurenzi IJ, Petersen PH, Zou K, Zhong W, Gerstein M, Snyder M. 2004 Major molecular differences between mammalian sexes are involved in drug metabolism and renal function. *Dev. Cell* **6**, 791–800. (doi:10.1016/j.devcel.2004.05.005)
- Vries GJ. 1990 Sex differences in neurotransmitter systems. *J. Neuroendocrinol.* **2**, 1–13. (doi:10.1111/j.1365-2826.1990.tb00385.x)
- Wolfe CA, Van Doren M, Walker HJ, Seney ML, McClellan KM, Tobet SA. 2005 Sex differences in the location of immunohistochemically defined cell populations in the mouse preoptic area/anterior hypothalamus. *Brain Res. Dev. Brain Res.* **157**, 34–41. (doi:10.1016/j.devbrainres.2005.03.001)
- Yang X, Schadt EE, Wang S, Wang H, Arnold AP, Ingram-Drake L, Drake TA, Lusis AJ. 2006 Tissue-specific expression and regulation of sexually dimorphic genes in mice. *Genome Res.* **16**, 995–1004. (doi:10.1101/gr.5217506)
- Veenig JG, Coolen LM. 1998 Neural activation following sexual behavior in the male and female rat brain. *Behav. Brain Res.* **92**, 181–193. (doi:10.1016/S0166-4328(97)00190-3)
- Coolen LM, Peters HJ, Veenig JG. 1997 Distribution of Fos immunoreactivity following mating versus anogenital investigation in the male rat brain. *Neuroscience* **77**, 1151–1161. (doi:10.1016/S0306-4522(96)00542-8)
- Gammie SC, Nelson RJ. 2001 cFOS and pCREB activation and maternal aggression in mice. *Brain Res.* **898**, 232–241. (doi:10.1016/S0006-8993(01)02189-8)
- Tobiansky DJ, Hattori T, Scott JM, Nutsch VL, Roma PG, Dominguez JM. 2012 Mating-relevant olfactory stimuli activate the rat brain in an age-dependent manner. *Neuroreport* **23**, 1077–1083. (doi:10.1097/WNR.0b013e32835b6ec1)
- Numan M, Numan MJ. 1997 Projection sites of medial preoptic area and ventral bed nucleus of the stria terminalis neurons that express Fos during maternal behavior in female rats. *J. Neuroendocrinol.* **9**, 369–384. (doi:10.1046/j.1365-2826.1997.t01-1-00597.x)
- Valcourt RJ, Sachs BD. 1979 Penile reflexes and copulatory behavior in male rats following lesions in the bed nucleus of the stria terminalis. *Brain Res. Bull.* **4**, 131–133. (doi:10.1016/0361-9230(79)90068-6)
- Liu YC, Salamone JD, Sachs BD. 1997 Lesions in medial preoptic area and bed nucleus of stria terminalis: differential effects on copulatory behavior and noncontact erection in male rats. *J. Neurosci.* **17**, 5245–5253.
- Emery DE, Sachs BD. 1976 Copulatory behavior in male rats with lesions in the bed nucleus of the

- stria terminalis. *Physiol. Behav.* **17**, 803–806. (doi:10.1016/0031-9384(76)90044-5)
27. Erb S, Stewart J. 1999 A role for the bed nucleus of the stria terminalis, but not the amygdala, in the effects of corticotropin-releasing factor on stress-induced reinstatement of cocaine seeking. *J. Neurosci.* **19**, 35.
28. Zardetto-Smith AM, Beltz TG, Johnson AK. 1994 Role of the central nucleus of the amygdala and bed nucleus of the stria terminalis in experimentally-induced salt appetite. *Brain Res.* **645**, 123–134. (doi:10.1016/0006-8993(94)91645-4)
29. Meloni EG, Jackson A, Gerety LP, Cohen BM, Carlezon WA Jr. 2006 Role of the bed nucleus of the stria terminalis (BST) in the expression of conditioned fear. *Ann. NY Acad. Sci.* **1071**, 538–541. (doi:10.1196/annals.1364.059)
30. Henke PG. 1984 The bed nucleus of the stria terminalis and immobilization-stress: unit activity, escape behaviour, and gastric pathology in rats. *Behav. Brain Res.* **11**, 35–45. (doi:10.1016/0166-4328(84)90006-8)
31. Schulz D, Canbeyli RS. 2000 Lesion of the bed nucleus of the stria terminalis enhances learned despair. *Brain Res. Bull.* **52**, 83–87. (doi:10.1016/S0361-9230(00)00235-5)
32. Sánchez MM, Aguado F, Sánchez-Toscano F, Saphier D. 1995 Effects of prolonged social isolation on responses of neurons in the bed nucleus of the stria terminalis, preoptic area, and hypothalamic paraventricular nucleus to stimulation of the medial amygdala. *Psychoneuroendocrinology* **20**, 525–541. (doi:10.1016/0306-4530(94)00083-M)
33. Hammack SE, Richey KJ, Watkins LR, Maier SF. 2004 Chemical lesion of the bed nucleus of the stria terminalis blocks the behavioral consequences of uncontrollable stress. *Behav. Neurosci.* **118**, 443–448. (doi:10.1037/0735-7044.118.2.443)
34. McHenry JA, Rubinow DR, Stuber GD. 2015 Maternally responsive neurons in the bed nucleus of the stria terminalis and medial preoptic area: putative circuits for regulating anxiety and reward. *Front. Neuroendocrinol.* **38**, 65–72. (doi:10.1016/j.yfrne.2015.04.001)
35. Gafford GM, Ressler KJ. 2015 GABA and NMDA receptors in CRF neurons have opposing effects in fear acquisition and anxiety in central amygdala vs. bed nucleus of the stria terminalis. *Horm. Behav.* **76**: 136–142. (doi:10.1016/j.yhbeh.2015.04.001)
36. Wu MV, Manoli DS, Fraser EJ, Coats JK, Tollkuhn J, Honda S, Harada N, Shah NM. 2009 Estrogen masculinizes neural pathways and sex-specific behaviors. *Cell* **139**, 61–71. (doi:10.1016/j.cell.2009.07.036)
37. Yang CF, Chiang MC, Gray DC, Prabhakaran M, Alvarado M, Juntti SA, Unger EK, Wells JA, Shah NM. 2013 Sexually dimorphic neurons in the ventromedial hypothalamus govern mating in both sexes and aggression in males. *Cell* **153**, 896–909. (doi:10.1016/j.cell.2013.04.017)
38. Unger EK, Burke Jr KJ, Yang CF, Bender KJ, Fuller PM, Shah NM. 2015 Medial amygdalar aromatase neurons regulate aggression in both sexes. *Cell Rep.* **10**, 453–462. (doi:10.1016/j.celrep.2014.12.040)
39. Juntti SA, Tollkuhn J, Wu MV, Fraser EJ, Soderborg T, Tan S, Honda SI, Harada, N, Shah NM. 2010 The androgen receptor governs the execution, but not programming, of male sexual and territorial behaviors. *Neuron* **66**, 260–272. (doi:10.1016/j.neuron.2010.03.024)
40. Shah NM, Pisapia DJ, Maniatis S, Mendelsohn MM, Nemes A, Axel R. 2004 Visualizing sexual dimorphism in the brain. *Neuron* **43**, 313–319. (doi:10.1016/j.neuron.2004.07.008)
41. Baum MJ. 2003 Activational and organizational effects of estradiol on male behavioral neuroendocrine function. *Scand. J. Psychol.* **44**, 213–220. (doi:10.1111/1467-9450.00338)
42. Crews D, Moore MC. 2005 Historical contributions of research on reptiles to behavioral neuroendocrinology. *Horm. Behav.* **48**, 384–394. (doi:10.1016/j.yhbeh.2005.04.003)
43. Dickson BJ. 2008 Wired for sex: the neurobiology of *Drosophila* mating decisions. *Science* **322**, 904–909. (doi:10.1126/science.1159276)
44. Haller J. 2013 The neurobiology of abnormal manifestations of aggression: a review of hypothalamic mechanisms in cats, rodents and humans. *Brain Res. Bull.* **93**, 97–109. (doi:10.1016/j.brainresbull.2012.10.003)
45. Manoli DS, Meissner GW, Baker BS. 2006 Blueprints for behavior: genetic specification of neural circuitry for innate behaviors. *Trends Neurosci.* **29**, 444–451. (doi:10.1016/j.tins.2006.06.006)
46. Moore FL, Boyd SK, Kelley DB. 2005 Historical perspective: hormonal regulation of behaviors in amphibians. *Horm. Behav.* **48**, 373–383. (doi:10.1016/j.yhbeh.2005.05.011)
47. Newman SW, Parfitt DB, Kollack-Walker S. 1997 Mating-induced c-fos expression patterns complement and supplement observations after lesions in the male Syrian hamster brain. *Ann. NY Acad. Sci.* **807**, 239–259. (doi:10.1111/j.1749-6632.1997.tb51924.x)
48. Perkins A, Roselli CE. 2007 The ram as a model for behavioral neuroendocrinology. *Horm. Behav.* **52**, 70–77. (doi:10.1016/j.yhbeh.2007.03.016)
49. Portman DS. 2007 Genetic control of sex differences in *C. elegans* neurobiology and behavior. *Adv. Genet.* **59**, 1–37. (doi:10.1016/S0065-2660(07)59001-2)
50. Siwicki KK, Kravitz EA. 2009 *Fruitless*, *doublesex* and the genetics of social behavior in *Drosophila melanogaster*. *Curr. Opin. Neurobiol.* **19**, 200–206. (doi:10.1016/j.conb.2009.04.001)
51. Torres CV, Sola RG, Pastor J, Pedrosa M, Navas M, García-Navarrete E, Ezquiaga E, García-Camba E. 2013 Long-term results of posteromedial hypothalamic deep brain stimulation for patients with resistant aggressiveness. *J. Neurosurg.* **119**, 277–287. (doi:10.3171/2013.4.jns121639)
52. Wade J, Arnold AP. 2004 Sexual differentiation of the zebra finch song system. *Ann. NY Acad. Sci.* **1016**, 540–559. (doi:10.1196/annals.1298.015)
53. Wallen K. 2005 Hormonal influences on sexually differentiated behavior in nonhuman primates. *Front. Neuroendocrinol.* **26**, 7–26. (doi:10.1016/j.yfrne.2005.02.001)
54. Fraser EJ, Shah NM. 2014 Complex chemosensory control of female reproductive behaviors. *PLoS ONE* **9**, e90368. (doi:10.1371/journal.pone.0090368)
55. Hasen NS, Gammie SC. 2009 *Trpc2* gene impacts on maternal aggression, accessory olfactory bulb anatomy and brain activity. *Genes Brain Behav.* **8**, 639–649. (doi:10.1111/j.1601-183X.2009.00511.x)
56. Hasen NS, Gammie SC. 2011 *Trpc2*-deficient lactating mice exhibit altered brain and behavioral responses to bedding stimuli. *Behav. Brain Res.* **217**, 347–353. (doi:10.1016/j.bbr.2010.11.002)
57. Kimchi T, Xu J, Dulac C. 2007 A functional circuit underlying male sexual behaviour in the female mouse brain. *Nature* **448**, 1009–1014. (doi:10.1038/nature06089)
58. Leybold BG, Yu CR, Leinders-Zufall T, Kim MM, Zufall F, Axel R. 2002 Altered sexual and social behaviors in *trp2* mutant mice. *Proc. Natl. Acad. Sci. USA* **99**, 6376–6381. (doi:10.1073/pnas.082127599)
59. Mandiyan VS, Coats JK, Shah NM. 2005 Deficits in sexual and aggressive behaviors in *Cnga2* mutant mice. *Nat. Neurosci.* **8**, 1660–1662. (doi:10.1038/nn1589)
60. Stowers L, Holy TE, Meister M, Dulac C, Koentges G. 2002 Loss of sex discrimination and male–male aggression in mice deficient for TRP2. *Science* **295**, 1493–1500. (doi:10.1126/science.1069259)
61. Wang Z, Balet Sindreu C, Li, V, Nudelman A, Chan GC-K, Storm DR. 2006 Pheromone detection in male mice depends on signaling through the type 3 adenylyl cyclase in the main olfactory epithelium. *J. Neurosci.* **26**, 7375–7379. (doi:10.1523/JNEUROSCI.1967-06.2006)
62. Wu Z, Autry AE, Bergan JF, Watabe-Uchida M, Dulac CG. 2014 Galanin neurons in the medial preoptic area govern parental behaviour. *Nature* **509**, 325–330. (doi:10.1038/nature13307)
63. Zufall F, Leinders-Zufall T. 2007 Mammalian pheromone sensing. *Curr. Opin. Neurobiol.* **17**, 483–489. (doi:10.1016/j.conb.2007.07.012)
64. Liberles SD. 2014 Mammalian pheromones. *Annu. Rev. Physiol.* **76**, 151–175. (doi:10.1146/annurevophys-021113-170334)
65. Touhara K, Vosshall LB. 2009 Sensing odorants and pheromones with chemosensory receptors. *Annu. Rev. Physiol.* **71**, 307–332. (doi:10.1146/annurev.physiol.010908.163209)
66. Dulac C, Wagner S. 2006 Genetic analysis of brain circuits underlying pheromone signaling. *Annu. Rev. Genet.* **40**, 449–467. (doi:10.1146/annurev.genet.39.07003.093937)
67. Wysocki CJ, Lepri JJ. 1991 Consequences of removing the vomeronasal organ. *J. Steroid Biochem. Mol. Biol.* **39**, 661–669. (doi:10.1016/0960-0760(91)90265-7)
68. Doudna JA, Charpentier E. 2014 Genome editing: The new frontier of genome engineering with

- CRISPR-Cas9. *Science* **346**, 1258096. (doi:10.1126/science.1258096)
69. Hsu PD, Lander ES, Zhang F. 2014 Development and applications of CRISPR-Cas9 for genome engineering. *Cell* **157**, 1262–1278. (doi:10.1016/j.cell.2014.05.010)
70. Sander JD, Joung JK. 2014 CRISPR-Cas systems for editing, regulating and targeting genomes. *Nat. Biotechnol.* **32**, 347–355. (doi:10.1038/nbt.2842)
71. Kim S, Ma L, Jensen KL, Kim MM, Bond CT, Adelman JP, Yu CR. 2012 Paradoxical contribution of SK3 and GIRK channels to the activation of mouse vomeronasal organ. *Nat. Neurosci.* **15**, 1236–1244. (doi:10.1038/nn.3173)
72. Omura M, Mombaerts P. 2014 Trpc2-expressing sensory neurons in the main olfactory epithelium of the mouse. *Cell Rep.* **8**, 583–595. (doi:10.1016/j.celrep.2014.06.010)
73. Edwards DA, Burge KG. 1971 Early androgen treatment and male and female sexual behavior in mice. *Horm. Behav.* **2**, 49–58. (doi:10.1016/0018-506X(71)90037-7)
74. Yoon H, Enquist LW, Dulac C. 2005 Olfactory inputs to hypothalamic neurons controlling reproduction and fertility. *Cell* **123**, 669–682. (doi:10.1016/j.cell.2005.08.039)
75. von Campenhausen H, Mori K. 2000 Convergence of segregated pheromonal pathways from the accessory olfactory bulb to the cortex in the mouse. *Eur. J. Neurosci.* **12**, 33–46. (doi:10.1046/j.1460-9568.2000.00879.x)
76. Scialfa F, Winans SS. 1975 The differential projections of the olfactory bulb and accessory olfactory bulb in mammals. *J. Comp. Neurol.* **161**, 31–55. (doi:10.1002/cne.901610105)
77. Winans SS, Scialfa F. 1970 Amygdaloid nucleus: new afferent input from the vomeronasal organ. *Science* **170**, 330–332. (doi:10.1126/science.170.3955.330)
78. Kang N, Baum MJ, Cherry JA. 2011 Different profiles of main and accessory olfactory bulb mitral/tufted cell projections revealed in mice using an anterograde tracer and a whole-mount, flattened cortex preparation. *Chem. Senses* **36**, 251–260. (doi:10.1093/chemse/bjq120)
79. Shipley MT, Adamek GD. 1984 The connections of the mouse olfactory bulb: a study using orthograde and retrograde transport of wheat germ agglutinin conjugated to horseradish peroxidase. *Brain Res. Bull.* **12**, 669–688. (doi:10.1016/036J.9230(84)90148-5)
80. Sosulski DL, Bloom ML, Cutforth T, Axel R, Datta SR. 2011 Distinct representations of olfactory information in different cortical centres. *Nature* **472**, 213–216. (doi:10.1038/nature09868)
81. Kvetter GA, Winans SS. 1981 Connections of the corticomедial amygdala in the golden hamster. I. Efferents of the ‘vomeronasal amygdala’. *J. Comp. Neurol.* **197**, 81–98. (doi:10.1002/cne.901970107)
82. Kvetter GA, Winans SS. 1981 Connections of the corticomedial amygdala in the golden hamster. II. Efferents of the ‘olfactory amygdala’. *J. Comp. Neurol.* **197**, 99–111. (doi:10.1002/cne.901970108)
83. Licht G, Meredith M. 1987 Convergence of main and accessory olfactory pathways onto single neurons in the hamster amygdala. *Exp. Brain Res.* **69**, 7–18. (doi:10.1007/BF00247024)
84. Meredith M. 1998 Vomeronasal, olfactory, hormonal convergence in the brain. Cooperation or coincidence? *Ann. NY Acad. Sci.* **855**, 349–361. (doi:10.1111/j.1749-6632.1998.tb10593.x)
85. Shipley MT, Murphy AZ, Rizvi TA, Ennis M, Behbehani MM. 1996 Olfaction and brainstem circuits of reproductive behavior in the rat. *Prog. Brain Res.* **107**, 355–377. (doi:10.1016/S0079-6123(08)61876-2)
86. Kang N, Baum MJ, Cherry JA. 2009 A direct main olfactory bulb projection to the ‘vomeronasal’ amygdala in female mice selectively responds to volatile pheromones from males. *Eur. J. Neurosci.* **29**, 624–634. (doi:10.1111/j.1460-9568.2009.06638.x)
87. Colpaert FC, Wiepkema PR. 1976 Effects of ventromedial hypothalamic lesions on spontaneous intraspecies aggression in male rats. *Behav. Biol.* **16**, 117–125. (doi:10.1016/S0009-6773(76)91225-6)
88. Commins D, Yahr P. 1984 Lesions of the sexually dimorphic area disrupt mating and marking in male gerbils. *Brain Res. Bull.* **13**, 185–193. (doi:10.1016/0361-9230(84)90020-0)
89. Goy RW, Phoenix CH. 1963 Hypothalamic regulation of female sexual behaviour; establishment of behavioural oestrus in spayed guinea-pigs following hypothalamic lesions. *J. Reprod. Fertil.* **5**, 23–40. (doi:10.1530/jrf.0.0050023)
90. Hennessy AC, Wallen K, Edwards DA. 1986 Preoptic lesions increase the display of lordosis by male rats. *Brain Res.* **370**, 21–28. (doi:10.1016/0006-8993(86)91100-5)
91. Kondo Y, Shinoda A, Yamanouchi K, Arai Y. 1990 Role of septum and preoptic area in regulating masculine and feminine sexual behavior in male rats. *Horm. Behav.* **24**, 421–434. (doi:10.1016/0018-506X(90)90019-T)
92. Kondo Y, Sachs BD, Sakuma Y. 1998 Importance of the medial amygdala in rat penile erection evoked by remote stimuli from estrous females. *Behav. Brain Res.* **91**, 215–222.
93. Kruk MR, van der Poel AM, de Vos-Frerichs TP. 1979 The induction of aggressive behaviour by electrical stimulation in the hypothalamus of male rats. *Behaviour* **70**, 292–322. (doi:10.1163/156853979X00106)
94. Lin D, Boyle MP, Dollar P, Lee H, Lein ES, Perona P, Anderson DJ. 2011 Functional identification of an aggression locus in the mouse hypothalamus. *Nature* **470**, 221–226. (doi:10.1038/nature09736)
95. Olivier B, Wiepkema PR. 1974 Behaviour changes in mice following electrolytic lesions in the median hypothalamus. *Brain Res.* **65**, 521–524. (doi:10.1016/0006-8993(74)90241-8)
96. Pfaff DW, Sakuma Y. 1979 Deficit in the lordosis reflex of female rats caused by lesions in the ventromedial nucleus of the hypothalamus. *J. Physiol.* **288**, 203–210. (doi:10.1113/jphysiol.1979.sp012691)
97. Pfaff DW, Sakuma Y. 1979 Facilitation of the lordosis reflex of female rats from the ventromedial nucleus of the hypothalamus. *J. Physiol.* **288**, 189–202. (doi:10.1113/jphysiol.1979.sp012690)
98. Yamanouchi K, Arai Y. 1985 Presence of a neural mechanism for the expression of female sexual behaviors in the male rat brain. *Neuroendocrinology* **40**, 393–397. (doi:10.1159/000124104)
99. Dugger BN, Morris JA, Jordan CL, Breedlove SM. 2007 Androgen receptors are required for full masculinization of the ventromedial hypothalamus (VMH) in rats. *Horm. Behav.* **51**, 195–201. (doi:10.1016/j.yhbeh.2006.10.001)
100. Guillamon A, Segovia S. 1997 Sex differences in the vomeronasal system. *Brain Res. Bull.* **44**, 377–382. (doi:10.1016/S0361-9230(97)00217-7)
101. Raisman G, Field PM. 1971 Sexual dimorphism in the preoptic area of the rat. *Science* **173**, 731–733. (doi:10.1126/science.173.3998.731)
102. Grgurevic N, Büdefeld T, Spanic T, Tobet SA, Majdic G. 2012 Evidence that sex chromosome genes affect sexual differentiation of female sexual behavior. *Horm. Behav.* **61**, 719–724. (doi:10.1016/j.yhbeh.2012.03.008)
103. Wersinger SR, Sannen K, Villalba C, Lubahn DB, Rissman EF, De Vries GJ. 1997 Masculine sexual behavior is disrupted in male and female mice lacking a functional estrogen receptor α gene. *Horm. Behav.* **32**, 176–183. (doi:10.1006/hbeh.1997.1419)
104. Zuloaga DG, Zuloaga KL, Hinds LR, Carbone DL, Handa RJ. 2014 Estrogen receptor β expression in the mouse forebrain: age and sex differences. *J. Comp. Neurol.* **522**, 358–371. (doi:10.1002/cne.23400)
105. Choi GB, Dong H-W, Murphy AJ, Valenzuela DM, Yancopoulos GD, Swanson LW, Anderson DJ. 2005 Lhx6 delineates a pathway mediating innate reproductive behaviors from the amygdala to the hypothalamus. *Neuron* **46**, 647–660. (doi:10.1016/j.neuron.2005.04.011)
106. Swanson LW. 2000 Cerebral hemisphere regulation of motivated behavior. *Brain Res.* **886**, 113–164. (doi:10.1016/S0006-8993(00)02905-X)
107. Juntti SA, Coats JK, Shah NM. 2008 A genetic approach to dissect sexually dimorphic behaviors. *Horm. Behav.* **53**, 627–637. (doi:10.1016/j.yhbeh.2007.12.012)
108. Sauer B. 1998 Inducible gene targeting in mice using the cre/lox system. *Methods* **14**, 381–392. (doi:10.1006/meth.1998.0593)
109. Branda CS, Dymecki SM. 2004 Talking about a revolution: the impact of site-specific recombinases on genetic analyses in mice. *Dev. Cell* **6**, 7–28. (doi:10.1016/S1534-5807(03)00399-X)
110. Chen SW, Shemyakin A, Wiedenmayer CP. 2006 The role of the amygdala and olfaction in unconditioned fear in developing rats. *J. Neurosci.* **26**, 233–240. (doi:10.1523/JNEUROSCI.2890-05.2006)
111. Cooke BM, Hegstrom CD, Keen A, Breedlove SM. 2001 Photoperiod and social cues influence the medial amygdala but not the bed nucleus of the stria terminalis in the Siberian hamster. *Neurosci.*

- Lett.* **312**, 9–12. (doi:10.1016/S0304-3940(01)02173-5)
112. Davis M. 1992 The role of the amygdala in fear and anxiety. *Annu. Rev. Neurosci.* **15**, 353–375. (doi:10.1146/annurev.ne.15.030192.002033)
113. Hong W, Kim DW, Anderson DJ. 2014 Antagonistic control of social versus repetitive self-grooming behaviors by separable amygdala neuronal subsets. *Cell* **158**, 1348–1361. (doi:10.1016/j.cell.2014.07.049)
114. Knapska E, Radwanska K, Werka T, Kaczmarek L. 2007 Functional internal complexity of amygdala: focus on gene activity mapping after behavioral training and drugs of abuse. *Physiol. Rev.* **87**, 1113–1173. (doi:10.1152/physrev.00037.2006)
115. Sah P, Faber ES, Lopez De Armentia M, Power J. 2003 The amygdaloid complex: anatomy and physiology. *Physiol. Rev.* **83**, 803–834. (doi:10.1152/physrev.00002.2003)
116. Swanson LW. 2003 The amygdala and its place in the cerebral hemisphere. *Ann. NY Acad. Sci.* **985**, 174–184. (doi:10.1111/j.1749-6632.2003.tb07081.x)
117. Bakker J, Honda S, Harada N, Balthazart J. 2003 The aromatase knockout (ArKO) mouse provides new evidence that estrogens are required for the development of the female brain. *Ann NY Acad Sci.* **1007**, 251–262. (doi:10.1196/annals.1286.024)
118. Toda K, Saibara T, Okada T, Onishi S, Shizuta Y. 2001 A loss of aggressive behaviour and its reinstatement by oestrogen in mice lacking the aromatase gene (*Cyp19*). *J. Endocrinol.* **168**, 217–220. (doi:10.1677/joe.0.1680217)
119. Harada N, Wakatsuki T, Aste N, Yoshimura N, Honda SI. 2009 Functional analysis of neurosteroidal oestrogen using gene-disrupted and transgenic mice. *J. Neuroendocrinol.* **21**, 365–369. (doi:10.1111/j.1365-2826.2009.01857.x)
120. Honda S, Harada N, Ito S, Takagi Y, Maeda S. 1998 Disruption of sexual behavior in male aromatase-deficient mice lacking exons 1 and 2 of the *cyp19* gene. *Biochem. Biophys. Res. Commun.* **252**, 445–449. (doi:10.1006/bbrc.1998.9672)
121. Morgan CW, Julien O, Unger EK, Shah NM, Wells JA. 2014 Turning on caspases with genetics and small molecules. *Methods Enzymol.* **544**, 179–213. (doi:10.1016/B978-0-12-417158-9.00008-X)
122. Sternson SM, Roth BL. 2014 Chemogenetic tools to interrogate brain functions. *Annu. Rev. Neurosci.* **37**, 387–407. (doi:10.1146/annurev-neuro-071013-014048)
123. Demas GE, Kriegsfeld LJ, Blackshaw S, Huang P, Gammie SC, Nelson RJ, Snyder SH. 1999 Elimination of aggressive behavior in male mice lacking endothelial nitric oxide synthase. *J. Neurosci.* **19**, RC30, 1–5.
124. Gammie SC, Nelson RJ. 1999 Maternal aggression is reduced in neuronal nitric oxide synthase-deficient mice. *J. Neurosci.* **19**, 8027–8035.
125. Svare B, Gandelman R. 1976 Suckling stimulation induces aggression in virgin female mice. *Nature* **260**, 606–608. (doi:10.1038/260606a0)
126. Baum MJ, Bakker J. 2013 Roles of sex and gonadal steroids in mammalian pheromonal communication. *Front. Neuroendocrinol.* **34**, 268–284. (doi:10.1016/j.yfrne.2013.07.004)
127. Bergan JF, Ben-Shaul Y, Dulac C. 2014 Sex-specific processing of social cues in the medial amygdala. *eLife* **3**, e02743. (doi:10.7554/eLife.02743)
128. DiBenedictis BT, Ingraham KL, Baum MJ, Cherry JA. 2012 Disruption of urinary odor preference and lordosis behavior in female mice given lesions of the medial amygdala. *Physiol. Behav.* **105**, 554–559. (doi:10.1016/j.physbeh.2011.09.014)
129. Sokolowski K, Corbin JG. 2012 Wired for behaviors: from development to function of innate limbic system circuitry. *Front. Mol. Neurosci.* **5**, 55. (doi:10.3389/fnmol.2012.00055)
130. Hess WR, Akert K. 1955 Experimental data on role of hypothalamus in mechanism of emotional behavior. *AMA Arch. Neurol. Psychiatry* **73**, 127–129. (doi:10.1001/archneurpsyc.1955.02330080005003)
131. Bard P. 1928 A diencephalic mechanism for the expression of rage with special reference to the sympathetic nervous system. *Am. J. Physiol.* **84**, 490–515.
132. Cheung CC, Krause WC, Edwards RH, Yang CF, Shah NM, Hnasko TS, Ingraham HA. 2015 Sex-dependent changes in metabolism and behavior, as well as reduced anxiety after eliminating ventromedial hypothalamus excitatory output. *Mol. Metab.* **4**, 857–866. (doi:10.1016/j.molmet.2015.09.001)
133. Hetherington AW, Ranson SW. 1940 Hypothalamic lesions and adiposity in the rat. *Anat. Rec.* **78**, 149–172. (doi:10.1002/ar.1090780203)
134. King BM. 2006 The rise, fall, and resurrection of the ventromedial hypothalamus in the regulation of feeding behavior and body weight. *Physiol. Behav.* **87**, 221–244. (doi:10.1016/j.physbeh.2005.10.007)
135. Kow LM, Harlan RE, Shivers BD, Pfaff DW. 1985 Inhibition of the lordosis reflex in rats by intrahypothalamic infusion of neural excitatory agents: evidence that the hypothalamus contains separate inhibitory and facilitatory elements. *Brain Res.* **341**, 26–34. (doi:10.1016/0006-8993(85)91468-4)
136. Kurrasch DM, Cheung CC, Lee FY, Tran PV, Hata K, Ingraham HA. 2007 The neonatal ventromedial hypothalamus transcriptome reveals novel markers with spatially distinct patterning. *J. Neurosci.* **27**, 13624–13634. (doi:10.1523/JNEUROSCI.2858-07.2007)
137. La Vaque TJ, Rodgers CH. 1975 Recovery of mating behavior in the female rat following VMH lesions. *Physiol. Behav.* **14**, 59–63. (doi:10.1016/0031-9384(75)90142-0)
138. Mathews D, Edwards DA. 1977 The ventromedial nucleus of the hypothalamus and the hormonal arousal of sexual behaviors in the female rat. *Horm. Behav.* **8**, 40–51. (doi:10.1016/0018-506X(77)90019-8)
139. Musatov S, Chen W, Pfaff DW, Kaplitt MG, Ogawa S. 2006 RNAi-mediated silencing of estrogen receptor α in the ventromedial nucleus of hypothalamus abolishes female sexual behaviors. *Proc. Natl. Acad. Sci. USA* **103**, 10 456–10 460. (doi:10.1073/pnas.0603045103)
140. Musatov S, Chen W, Pfaff DW, Mobbs CV, Yang X-J, Clegg DJ, Kaplitt MG, Ogawa S. 2007 Silencing of estrogen receptor α in the ventromedial nucleus of hypothalamus leads to metabolic syndrome. *Proc. Natl. Acad. Sci. USA* **104**, 2501–2506. (doi:10.1073/pnas.0610787104)
141. Reeves AG, Plum F. 1969 Hyperphagia, rage, and dementia accompanying a ventromedial hypothalamic neoplasm. *Arch. Neurol.* **20**, 616–624. (doi:10.1001/archneur.1969.00480120062005)
142. Robarts DW, Baum MJ. 2007 Ventromedial hypothalamic nucleus lesions disrupt olfactory mate recognition and receptivity in female ferrets. *Horm. Behav.* **51**, 104–113. (doi:10.1016/j.yhbeh.2006.08.009)
143. Silva BA, Mattucci C, Krzywkowski P, Murana E, Illarionova A, Grinevich V, Canteras NS, Ragozzino D, Gross CT. 2013 Independent hypothalamic circuits for social and predator fear. *Nat. Neurosci.* **16**, 1731–1733. (doi:10.1038/nn.3573)
144. Swaab D. 2003 Chapter 9 The ventromedial nucleus (VMN; nucleus of Cajal). *Handb. Clin. Neurol.* **79**, 239–242. (doi:10.1016/S0072-9752(03)80016-7)
145. Spiteri T, Musatov S, Ogawa S, Ribeiro A, Pfaff DW, Agmo A. 2010 Estrogen-induced sexual incentive motivation, proceptivity and receptivity depend on a functional estrogen receptor α in the ventromedial nucleus of the hypothalamus but not in the amygdala. *Neuroendocrinology* **91**, 142–154. (doi:10.1159/000255766)
146. Spiteri T, Musatov S, Ogawa S, Ribeiro A, Pfaff DW, Agmo A. 2010 The role of the estrogen receptor α in the medial amygdala and ventromedial nucleus of the hypothalamus in social recognition, anxiety and aggression. *Behav. Brain Res.* **210**, 211–220. (doi:10.1016/j.bbr.2010.02.033)
147. Falkner AL, Dollar P, Perona P, Anderson DJ, Lin D. 2014 Decoding ventromedial hypothalamic neural activity during male mouse aggression. *J. Neurosci.* **34**, 5971–5984. (doi:10.1523/JNEUROSCI.5109-13.2014)
148. Lee H, Kim DW, Remedios R, Anthony TE, Chang A, Madisen L, Zeng H, Anderson DJ. 2014 Scalable control of mounting and attack by ESR1 $^{+}$ neurons in the ventromedial hypothalamus. *Nature* **509**, 627–632. (doi:10.1038/nature13169)
149. Lo L, Anderson DJ. 2011 A Cre-dependent, anterograde transsynaptic viral tracer for mapping output pathways of genetically marked neurons. *Neuron* **72**, 938–950. (doi:10.1016/j.neuron.2011.12.002)
150. Luo L, Callaway EM, Svoboda K. 2008 Genetic dissection of neural circuits. *Neuron* **57**, 634–660. (doi:10.1016/j.neuron.2008.01.002)
151. Petreanu L, Huber D, Sobczyk A, Svoboda K. 2007 Channelrhodopsin-2-assisted circuit mapping of long-range callosal projections. *Nat. Neurosci.* **10**, 663–668. (doi:10.1038/nn1891)
152. Wall NR, Wickersham IR, Cetin A, De La Parra M, Callaway EM. 2010 Monosynaptic circuit tracing *in vivo* through Cre-dependent targeting and complementation of modified rabies virus. *Proc. Natl. Acad. Sci. USA* **107**, 21 848–21 853. (doi:10.1073/pnas.1011756107)

153. Wickersham IR, Lyon DC, Barnard RJ, Mori T, Finke S, Conzelmann KK, Young JA, Callaway EM. 2007 Monosynaptic restriction of transsynaptic tracing from single, genetically targeted neurons. *Neuron* **53**, 639–647. (doi:10.1016/j.neuron.2007.01.033)
154. Akerboom J *et al.* 2012 Optimization of a GCaMP calcium indicator for neural activity imaging. *J. Neurosci.* **32**, 13 819–13 840. (doi:10.1523/JNEUROSCI.2601-12.2012)
155. Chen TW *et al.* 2013 Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature* **499**, 295–300. (doi:10.1038/nature12354)
156. Cui G, Jun SB, Jin X, Pham MD, Vogel SS, Lovinger DM, Costa RM. 2013 Concurrent activation of striatal direct and indirect pathways during action initiation. *Nature* **494**, 238–242. (doi:10.1038/nature11846)
157. Chen Y, Lin YC, Kuo TW, Knight ZA. 2015 Sensory detection of food rapidly modulates arcuate feeding circuits. *Cell* **160**, 829–841. (doi:10.1016/j.cell.2015.01.033)
158. Gunaydin LA *et al.* 2014 Natural neural projection dynamics underlying social behavior. *Cell* **157**, 1535–1551. (doi:10.1016/j.cell.2014.05.017)
159. Hamel EJ, Grewe BF, Parker JG, Schnitzer MJ. 2015 Cellular level brain imaging in behaving mammals: an engineering approach. *Neuron* **86**, 140–159. (doi:10.1016/j.neuron.2015.03.055)
160. Raskin K, de Gendt K, Duittoz A, Liere P, Verhoeven G, Tronche F, Mhaouty-Kodja S. 2009 Conditional inactivation of androgen receptor gene in the nervous system: effects on male behavioral and neuroendocrine responses. *J. Neurosci.* **29**, 4461–4470. (doi:10.1523/JNEUROSCI.0296-09.2009)
161. Bangasser DA, Valentino RJ. 2014 Sex differences in stress-related psychiatric disorders: neurobiological perspectives. *Front. Neuroendocrinol.* **35**, 303–319. (doi:10.1016/j.yfrne.2014.03.008)
162. Davies W. 2014 Sex differences in Attention Deficit Hyperactivity Disorder: candidate genetic and endocrine mechanisms. *Front. Neuroendocrinol.* **35**, 331–346. (doi:10.1016/j.yfrne.2014.03.003)
163. Li R, Singh M. 2014 Sex differences in cognitive impairment and Alzheimer's disease. *Front. Neuroendocrinol.* **35**, 385–403. (doi:10.1016/j.yfrne.2014.01.002)
164. Smith KM, Dahodwala N. 2014 Sex differences in Parkinson's disease and other movement disorders. *Exp. Neurol.* **259**, 44–56. (doi:10.1016/j.expneurol.2014.03.010)
165. Werling DM, Geschwind DH. 2013 Sex differences in autism spectrum disorders. *Curr. Opin. Neural.* **26**, 146–153. (doi:10.1097/WCO.0b013e32835ee548)